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Abstract

Utilization of biomass as a new and renewable energy source is being actively conducted by various parties. One of the technology for utilizing or converting biomass as an energy source is pyrolysis, to convert biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from vapors phase of biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. The total phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). The content of phenol compounds in corrosive bio-oils still quite high, the use of this bio-oil directly will cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive, and unstable. Phenol compounds have some benefits as one of the compound for floor cleaner and disinfectant which contained in bio-oil. The correlation between experimental data and calculations shows that the UNIQUAC Functional-group Activity Coefficients (UNIFAC) equilibrium model can be used to predict the liquid-liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the Non Random Two Liquid (NRTL) equilibrium model can be used to predict liquid-liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells.

Keywords	biomass, pyrolysis, bio-oil, UNIFAC, NRTL
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Liquid Phase Equilibrium of Phenol Extraction from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models

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ABSTRACT

Utilization of biomass as a new and renewable energy source is being actively conducted by various parties. One of the technology for utilizing or converting biomass as an energy source is pyrolysis, to convert biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from vapors phase of biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. Bio-oil from coffee shells contains 34% phenol, while 40.01% as for phenol content of coconut shell. The content of phenol compounds in corrosive bio-oils still quite high, causes the use of this bio-oil directly will cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive, and unstable. Phenol compounds have some benefits as one of the compound for floor cleaner and disinfectant which contained in bio-oil. The correlation between experimental data and calculations shows that the UNIFAC equilibrium model can be used to predict the liquid-liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the NRTL equilibrium model can be used to predict liquid-liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells.

Keywords: biomass, pyrolysis, bio-oil, UNIFAC, NRTL.

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1. INTRODUCTION

The role of fossil fuels still be crucial, but their effects will gradually be over taken by new and renewable energy sources. Along with the depletion of world oil reserves and environmental issues, various researches on renewable energy are being actively conducted by various parties. Therefore, technology needs be able to overcome the problem. One technology that can be used is pyrolysis technology where the material used comes from organic waste (biomass). Pyrolysis technology can overcome organic waste problems and make the environment healthy also convert it into value-added end product.

Biomass is an organic material derived from plants, algae, animal waste, industrial waste, forestry, and agricultural waste [1]. Biomass can be used as pyrolysis raw material to produce liquid (bio-oil), gas and solid products. Various kinds of biomass waste can be used as raw materials for the manufacture of bio-oils, such as corn cobs [2], sawdust and wheat bran [3], castor [4], oak [5], durian shell [6] and coconut shell [7].

Indonesia is the third largest coffee producer in the world after Brazil and Vietnam by contributing about 6% of the world's total coffee production, also world's fourth largest coffee exporter with a market share of around 11% across the world. Coffee is one of the plantations with high economic value besides chocolate and tea. Based on data from Indonesian Ministry of Forestry and Plantation, Indonesia's coffee production in 2013 reached 691,160 tons with a plantation area of 47,800 Ha [8]. Waste from coffee processing that abundant and not utilized optimally is the coffee shell. Large waste of coffee shells derived from plantations was approximately 11.08% which potentially cause environment hazard. Coffee shells are a waste that has high lignocellulose content which have potential to be used as bio-oil. Bio-oil is a blackish liquid fuel derived from biomass such as wood, coffee shells and other biomass. Bio-oil contains lignin components (23.4%), cellulose (34.2%) and hemicellulose (24.5%) [9].

In Indonesia, in addition to the shell, one of the available biomass resources is coconut shell. Availability of coconut shell can be seen from the production of coconut fruit in Indonesia. Coconut is one of Indonesia's main commodities with an average production of 3,000,000 tons / year [10]. The utilization of coconut is very diverse, but for the utilization and way of handling waste is still limited. Coconut fruit can be used as food in daily life for the purpose of consumed directly. Coconut shells nowadays, lots of it become garbage that can not be utilized. One of the waste handling is to utilize coconut shell waste as a base material in the manufacture of bio-oil. Characterization studies on coconut shells have been done through proximate, ultimate, and thermo gravimetric (TG) analyzes. The results of this analysis indicate that coconut shell has a high potential to produce fuel fluid by pyrolysis conversion process [11]. Chemical content of coconut shell is cellulose (34%), hemicellulose (21%), lignin (27%) and ash (18%) [12].

Before converted into bio-oil, coconut shell waste and coffee shell need to be treated first. It takes a process

before converting the waste of coffee shell into bio-oil which is pyrolysis. Pyrolysis is the process of decomposition of a substance or material that is done at a relatively high temperature. Pyrolysis results in the form of gas, liquid and solid. The solid result of this process is char (char). Gas and liquid products can be used as a source of bio-oil. Bio-oil can be used as an alternative fuel that is easier to store and distribute than biomass in solid form. Bio-oil made from coconut shell material using slow pyrolysis with pyrolysis temperature 250-300 °C produces liquid which very acidic and corrosive. The largest organic component in bio-oil is the lignin derivative of phenol, alcohol, organic acids and carbonyl compounds such as ketones, aldehydes and esters [13]. The nature of the acid is caused by the content of the dominant phenol compounds.

Phenol compounds in addition to causing the smell of smoke in bio-oil, also showed antioxidant activity and corrosive [14]. Therefore, the use of this bio-oil will directly cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive and unstable [6].

Phenol needs to be extracted from bio-oil to reduce its corrosive properties. In addition, phenol is a compound that has a high economic value because of its expensive and widely used. Phenols have many important uses such as disinfectants, resins, pesticides, explosives, and drugs. The method that can be used to extract phenol compounds from bio-oil is by liquid-liquid extraction method using methanol solvent. This has been done by Jazbinsk [15] on the isolation of phenol compounds from bio-oils from pyrolysis of forest waste and Manilla [16] on the extraction of phenol compounds from bio-oils from pyrolysis of agricultural waste. Dahlia [17] conducted research on liquid-liquid extraction of phenol compound from bio-oil of pyrolysis resulted of palm oil empty bunch using methanol solvent. The yield of phenol resulting from this extraction is 40%. In this study, it is expected that the liquid-liquid extraction method can be used to separate the phenol compounds from coconut shell pyrolysis bio-oils. The purpose of phenol extraction is to improve the quality of bio-oil and reduce the corrosivity, so when used as fuel, it will cause no damage to the engine.

Biomass pyrolysis converted to bio-oil studies have been conducted. Bio-oil produced from the pyrolysis process can be used as fuel for engines, turbines and boilers. The bio-oil which processed further by liquid-liquid extraction process will reduce the corrosive nature of bio-oil and obtained chemicals which one of them is phenol. Research on extraction of phenol compounds from bio-oil is still very limited. In addition, thermodynamic modelling study on liquid-liquid equilibrium of phenol extraction process from bio-oil still needs to be investigated further.

To design extraction columns required various information such as liquid-liquid equilibrium data (Liquid-Liquid Equilibrium). Thermodynamic models such as Equation of State or Activity Coefficient are used to correlate the experimental data to obtain optimal interaction parameters. Optimal parameters can be used to predict liquid-liquid equilibrium data to produce an equilibrium graph that is used to design extraction columns [18]. The study on liquid-liquid equilibrium include NRTL, e-NRTL, UNIQUAC, and UNIFAC.

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Given the importance of thermodynamic modeling in this liquid-liquid extraction process and the price and benefits of phenol in the industrial world, this research becomes an important thing to do. From the equilibrium data, the optimum condition will be obtained when the highest yield of extract obtained in the extraction process of phenol compound from bio-oil from coconut pyrolysis. Liquid-liquid equilibrium data of the components of the system can be used in the design of the equipment in separation process.

2. EXPERIMENTAL SECTION

2.1 Chemicals

All reagents used here are A.R. grade and used as received without further purification. Methanol, acetic acid, and chloroform were purchased from e-Merck (Germany). The distilled water is available at the laboratory. Coffee shell and coconut shell were obtained from public market in Semarang.

2.2 Equilibrium Procedure

Instruments used in this study:

- Shaker bath, product by Memmert WB14, SV 1422, Schwabach, Germany.
- Gas Chromatography (GC), product by GC 6820 Agilent Technologies, column: Rastek RXi-5MS002E.
- Gas Chromatography-Mass Spectrometry (GC-MS), product by GCMS-QP2010S Shimadzu, column: Rastek RXi-5MS.

The material systems and experimental conditions for the equilibrium extraction are summarized in Table 1.

Table 1. Material Systems and Conditions for Equilibrium Extraction

The bio-oil feed solution that containing phenol was extracted using two-stage extraction. The first stage uses aquadest as a polar solvent and chloroform as a non-polar solvent. Extraction was conducted with ice-bath method at 4-5°C, 200 rpm and stirring time for 2 hours. The extracts obtained at this first stage are used as feeds on the second stage extraction. At the second stage extraction, each extract was extracted using a methanol solvent under operating conditions shown in Table 1. The extraction equipment set can be seen in Figure 1.

Figure 1. Extraction Equipment

2.3 Analysis and Characterization

After equilibrium attained (60 minutes), the mixtures were poured into a separating funnel, settled for an hour and separated into two phases. Then the two phases were weighed. The extract phase and the raffinate phase of each extraction processes are analyzed by GC to obtain the concentration of each component. The principal conditions of this analysis are shown in Table 2.

Table 2. Conditions of Analysis Using GC

3. RESULTS AND DISCUSSION

3.1 Bio-Oil Content

Gas Chromatography-Mass Spectrometry (GC-MS) analysis used to determine total content of phenol in coconut shell bio-oil. It shows that coconut shell bio-oil contained total phenol for 40.01%. The results of GC-MS bio-oil analysis of coconut shell pyrolysis results can be seen in Figures 2 and 3.

Figure 2. Analysis of GC-MS from Coconut shell Bio-oil

Table 3. Component of Bio-oil from Coconut shell

Figure 3. Analysis of GC-MS from Coffee Shell Bio-oil

Table 4. Component of Bio-oil from Coffee Shell

Figure 2 and 3, Table 3 and 4 represent that bio-oil contains more than 5 main components. The percentage (%) of the compositions of the bio-oil analysis by GC-MS was obtained from the percentage area of the peak or the height of the peak in the chromatograms. Using a method of GC-MS, this work presents a composition of the bio-oil generated in the experiment. The total phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). It was caused by the pyrolysis processing at 300-350°C (low temperature carbonization) that produce more substances contain ester and phenolic compounds. The composition of biomass such as hemicellulose, cellulose and lignin will be oxidized to phenol as the main component of the bio-oil.

3.2 Equilibrium data on phenol extraction of bio-oil from coconut pyrolysis results

In this research the variables studied were temperatur, speed of stirring and type of solvents (aqueous methanol and aqueous acetic acid). Generally, all extracts are rich of phenol but the mole fraction significantly depends on extraction conditions: temperatur, speed of stirring and type of solvents. Methanol and acetic acid need to be mixed with water because their polarities are not strong enough to form immiscible mixture with bio-oil [19].

The mole fractions of any components in the raffinate phase and the extract phase, x_i and y_i , are constant after 60 minutes, so the system is assumed to be in equilibrium. The amount of water transferred from the solvent into the raffinate phase was negligible [19].

In this work, we are reported experimental data for the phenol extraction from bio-oil produced from tempurung kelapa and the phenol extraction from bio-oil produced from kangkang kopi.

Table 5 and 6 show the equilibrium of single-components extraction data for aqueous acetone and aqueous methanol solvent in the extract phase and raffinate phase.

Table 5. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Methanol from Bio-oil Produced from Coconut Shell.

Table 6. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Acetic Acid from Bio-oil Produced from Coffee Shell.

3.3 Liquid-liquid Equilibrium for The Phenol Extraction Process using UNIFAC dan NRTL Models.

The UNIFAC and NRTL models were used to correlate the experimental data obtained for the systems under study. UNIFAC equation and NRTL equation (Smith, 1996) were used to correlate experimental phase equilibrium data. The activity coefficient models applied in this study are as follows:

a) UNIFAC: [20]

UNIFAC is a UNIQUAC Functional-group Activity Coefficients model based on the UNIQUAC equation by using functional groups of a molecule contained in the mixture to calculate the activity coefficients. This model was developed by Fredenslund, Jones, and Prausnitz (1975). The UNIFAC modeling is a continuation of the UNIQUAC modeling development. This model has the advantage of performing phase equilibrium estimation even in the absence of experimental data. This thermodynamic model is used to predict the estimation of non-electrolyte activity in non-ideal mixtures. The UNIFAC model calculates the activity coefficients by using functional groups present in the molecules that form the liquid mixture. If the value of the component activity coefficient has a value close to one, this indicates that the component is in the ideal state in the system [21]. The correlation in the UNIFAC model is used in order to reduce the number of binary interactions measured to predict the state of the system. The correlation for the UNIFAC model is shown as follows:

$$\ln \gamma_i = \ln \gamma_i^c + \ln \gamma_i^R \quad (1)$$

$$\ln \gamma_i^c = 1 - J_i + \ln J_i - 5q_i \left(1 - \frac{J_i}{L_i} + \ln \frac{J_i}{L_i} \right) \quad (2)$$

$$\ln \gamma_i^R = q_i \left[1 - \sum_k \left(\theta_{ik} \frac{R_k}{R_i} - e_{ik} \ln \frac{R_k}{R_i} \right) \right] \quad (3)$$

$$J_i = \frac{r_i q_i}{L_i} \quad (4)$$

$$L_i = \frac{q_i r_i}{\sum_j q_j r_j} \quad (5)$$

$$r_i = \sum_k v_k^{(i)} R_k \quad (6)$$

$$q_i = \sum_k v_k^{(i)} Q_k \quad (7)$$

$$e_{ik} = \frac{u_{ik}^{(i)}}{q_i} \quad (8)$$

$$\beta_{ik} = \sum_m e_{im} r_m \quad (9)$$

$$\theta_{ik} = \frac{\sum_j r_j q_{ij}}{\sum_j r_j q_j} \quad (10)$$

$$s_k = \sum_m \beta_m r_m \quad (11)$$

$$\tau_{mk} = \exp \left(-\frac{u_{mk}}{T} \right) \quad (12)$$

Where:

- $\ln \gamma_i^c$ – Combinatorial component
- $\ln \gamma_i^R$ – residual component
- J_i – component surface area component (i)
- L_i – component volume fraction (i)
- r_i – pure component surface area parameter (i)
- x_i – pure component mol fraction (i)
- q_i – pure component volume parameter (i)
- $v_k^{(i)}$ – number of sub-group (k) on component (i)
- R_k – parameter area for sub-group (k) ($R_k = A_{sk}/(2.5 \times 10^8)$), where A_{sk} is number of sub-group van der Waals surface area
- Q_k – number of parameter volume subgroup (k) ($Q_k = V_{sk}/15.17$, where V_{sk} is number of Van der Waals volume subgroup)
- u_{ik} – group of interaction parameter from group m and k [18]
- x_i – liquid mol fraction of component (i)

UNIFAC equation can be obtained to :

$$\ln \gamma_i = \ln \left[1 - J_i + \ln J_i + 5q_i \left(1 - \frac{J_i}{L_i} + \ln \frac{J_i}{L_i} \right) \right] + \ln \left[1 - \sum_k \left(\frac{\sum_j r_j q_{ij}}{\sum_j r_j q_j} \frac{R_k}{R_i} - \frac{u_{ik}^{(i)}}{q_i} \ln \frac{\sum_j r_j q_{ij}}{\sum_j r_j q_j} \right) \right] \quad (13)$$

b) NRTL: [20]

Non Random Two Liquid (NRTL) is derived from the Scott two liquid model and it is assumed that randomness is the same as the model used on the Wilson model. The NRTL model has a non-random parameter (α_{12}) making it possible to use in a wide variety of mixtures. The NRTL model uses only binary parameters to calculate the phase equilibrium properties.

The NRTL (Non Random Two-Liquid) equation was first developed by Renon and Prausnitz (1968). The NRTL model is derived from the Scott two liquid model and it is assumed that the non-random parameter is the same as the model used in the Wilson Model. The NRTL equation is developed based on a two-fluid theory

with the addition of a non-random parameter (α), which makes the equation applicable to different types of liquid-liquid mixture and equilibrium by choosing the appropriate α value. The parameter estimation for this equation is done by setting the α value at a certain value and subsequently optimizing the objective function to obtain two optimal parameter interactions. The recommended α value is 0.2 - 0.47. The value of α is often adjusted to certain conditions, but usually the value of $\alpha = 0.3$. Unlike the Wilson equation which contains two adjustable parameters, the NRTL equation contains three parameters, so the NRTL equation can be an alternative model if other models are inadequate. This equation can also estimate the equilibrium of multicomponent systems without additional parameters.

The NRTL equation can represent liquid-liquid equilibrium for both binary and multicomponent systems only with binary parameters. The equation is simpler than the UNIQUAC equation, but it has disadvantage because it contains three parameters for each pair of components. The third parameter (α) is estimated based on the state of the component and sometimes for certain substances the extreme number is obtained. The NRTL equation is expressed:

$$\frac{G^E}{RT} = \sum_{i=1}^n \sum_{j=1}^n x_i x_j \frac{g_{ij} \tau_{ij}}{z_{ij} + \alpha_{ij} g_{ij}} \quad (14)$$

$$\tau_{ij} = \frac{g_{ij} - g_{ji}}{RT} \quad (15)$$

$$g_{ii} = g_{jj} = g_{kk} = 0 \quad (16)$$

$$g_{ij} = \exp(-\alpha_{ij} \tau_{ij}) \quad (17)$$

$$\alpha_{ij} = \alpha_{ji} \quad (18)$$

$$\ln y_i = \frac{\sum_{j=1}^n x_j \tau_{ij} g_{ij}}{\sum_{k=1}^n x_k \tau_{ik} g_{ik}} + \sum_{j=1}^n x_j \frac{g_{ij}}{z_{ij} + \alpha_{ij} g_{ij}} - \sum_{k=1}^n \frac{x_j \tau_{jk} g_{jk}}{z_{jk} + \alpha_{jk} g_{jk}} \quad (19)$$

$$X_i = \exp \left(\frac{\sum_{j=1}^n x_j \tau_{ij} g_{ij}}{\sum_{k=1}^n x_k \tau_{ik} g_{ik}} + \sum_{j=1}^n x_j \frac{g_{ij}}{z_{ij} + \alpha_{ij} g_{ij}} - \sum_{k=1}^n \frac{x_j \tau_{jk} g_{jk}}{z_{jk} + \alpha_{jk} g_{jk}} \right) \quad (20)$$

$$Y_j = \exp \left(\frac{\sum_{i=1}^n x_i \tau_{ij} g_{ij}}{\sum_{k=1}^n x_k \tau_{ik} g_{ik}} + \sum_{i=1}^n x_i \frac{g_{ij}}{z_{ij} + \alpha_{ij} g_{ij}} - \sum_{k=1}^n \frac{x_i \tau_{ki} g_{ki}}{z_{ki} + \alpha_{ki} g_{ki}} \right) \quad (21)$$

Where:

- G^E = molar Gibbs energy
- R = gas constant
- T = temperature
- x = liquid phase mol fraction
- i, j, k = component of 1, 2, 3, ..., n
- α = non-random parameter for NRTL.

- g, G = component parameter
- γ = activity coefficient
- τ = empirical constant for NRTL.

Table 7. The Activity Coefficients were Calculated by UNIFAC Model.

Table 8. The Activity Coefficients were Calculated by NRTL Model.

Table 7 and 8 shows the value of activity coefficient on phenol extraction using methanol solvent obtained from calculation with UNIFAC and NRTL modeling, from coefficient value of activity on each component shows that methanol and acetic acid component in extract phase is in ideal condition because the value of activity coefficient obtained close to one. The activity coefficient value of phenol and water activity in the extract phase is not close to one, it means that the phenol and water components are in less than ideal conditions in the system. The ideal conditions of the components in the system show that the component is dominant, so the properties of the mixture are closer to the dominant component properties either in the extract or raffinate phase. In raffinate it is known that the solvent has a coefficient of activity close to the one, indicating that the solvent is in ideal conditions in the system, while the phenol and bio-oil components are in less than ideal conditions. So non-ideality studies are very important for those systems [21].

Figure 4. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 298 K.

Where:

- Xa: mole fraction of component in extract phase
- Xb: mole fraction of solvent in extract phase
- Xc: mole fraction of water in extract phase
- Ya: mole fraction of component in raffinate phase
- Yb: mole fraction of solvent in raffinate phase
- Yd: mole fraction of bio-oil in raffinate phase

Figure 5. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 313 K.

Figure 6. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 323 K.

Figure 7. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 298 K.

Figure 8. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 313 K.

Figure 9. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 323 K.

Figure 10. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 298 K.

Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 313 K.

Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 323 K.

Figure 13. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 298 K.

Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 313 K.

Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Acetic Acid by NRTL Model at 323 K.

The result of correlation of liquid-liquid equilibrium data by using UNIFAC and NRTL modeling gives good agreement between experimental data and calculation. This is shown by the value of SSE (Sum of the Square of the Error) approaching the value of 0. SSE values in UNIFAC and NRTL modeling can be seen in Table 8.

Table 8. Sum of the Square of the Error (SSE)

Based on Table 8, it can be seen that extraction with methanol solvent with temperature of 323 K has the smallest SSE value compared to other conditions with SSE value of 0.053 (for UNIFAC model) and 0.015 (for NRTL model). The smaller the SSE values obtained show that the correlation between the experimental data and the calculation data indicates a better correlation.

Good correlation results show that equilibrium modeling of UNIFAC and NRTL can be used to predict liquid-liquid equilibrium in phenol extraction process from coconut shell pyrolysis bio-oil and coffee shell using methanol solvent and acetic acid at 323 K.

4. CONCLUSION

The UNIFAC modeling provides a good correlation to liquid-liquid equilibrium data for ternary systems on phenol extraction of coconut shell pyrolysis bio-oil using methanol solvent under conditions of temperature of 323 K given by SSE of 0.053.

NRTL modeling gives a good correlation to liquid-liquid equilibrium data for ternary system on phenol extraction of bio-oil from pyrolysis of coffee shell using methanol solvent at temperature condition of 323 K given by SSE of 0.015.

The activity coefficients of methanol, acetic acid, and phenol in the extract phase and raffinate phase are far from unity. It means that they are far from ideal condition. The activity coefficients of water and kerosene in their phases are close to one. It means that they are nearly ideal in their phases.

ACKNOWLEDGMENTS

The authors would like to acknowledge the Ministry of Research, Technology and Higher Education, Indonesia, for financial support of this work through the research grant of "Produk Terapan" Universitas Negeri Semarang, Nomor: 084/SP2/H/TL/DRPM/IV/2017.

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Table 1. Material Systems and Conditions for Equilibrium Extraction.

	Quantity
feed	bio-oil
feed volume	2.5 mL
solvent	aqueous methanol aqueous acetic acid
mole fraction of water in solvent	0.2
solvent-feed mass ratio	1
shaking time	60 minutes
amplitude of shaking	150, 200, 250 rpm
temperature	298, 313, 323K.

Table 2. Conditions of Analysis Using GC

Column: Rastek RXG-SMS		
Column:		
Inner diameter	[m]	3.2 x 10 ⁻⁴
Length	[m]	30
Carrier gas		He
Split ratio	[-]	153
Flow rate	[cm ³ /sec]	26.6
Sample volume	m ³	1.10 ⁻⁹
Injection temperature	[K]	553
Column temperature	[K]	313 – 573
Pressure column	kPa	10.0
Column flow	mL/min	0.54
Detector (FID) temperature	[K]	573

13

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Table 3. The Main Components of Bio-oil Produced by Pyrolysis of Coconut Shells

Peak	Component	Percentage
1	Ethyl ester	37.60
2	Formic acid	4.02
3	1-Hydroxy-2-butanone	3.89
4	Furfural	5.45
5	Phenol	40.01
7	2-Methoxy-phenol	7.02

Table 4. The Main Components of Bio-oil Produced by Pyrolysis of Coffee Shells

Peak	Component	Percentage
1	Urea	5%
2	Acetic acid	26%
5	Furfural	6%
6	Phenol	34%
7	Phenol, 2-methoxy-	7%
8	Phenol, 2,6-dimethoxy	4%

Table 5. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Methanol from Bio-oil Produced from Coconut Shell

Temperature	Speed	X _a	X _b	X _c	Y _a	Y _b	Y _d
298 K	150	0.007	0.973	0.020	0.161	0.316	0.524
	200	0.007	0.973	0.019	0.116	0.313	0.572
	250	0.009	0.967	0.024	0.094	0.488	0.418
313 K	150	0.009	0.963	0.027	0.085	0.549	0.365
	200	0.008	0.970	0.022	0.086	0.414	0.501
	250	0.010	0.963	0.027	0.062	0.555	0.383
323 K	150	0.010	0.962	0.028	0.069	0.567	0.363
	200	0.009	0.965	0.026	0.065	0.525	0.410
	250	0.012	0.959	0.029	0.029	0.615	0.356

Table 6. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Acetic Acid from

Bio-oil Produced from Coffee Shell.

Temperature	Speed	X _a	X _b	X _c	Y _a	Y _b	Y _d
298 K	150	0.054	0.247	0.365	0.281	0.047	0.672
	200	0.042	0.370	0.204	0.107	0.537	0.357
	250	0.024	0.551	0.093	0.019	0.878	0.104
313 K	150	0.012	0.707	0.068	0.025	0.907	0.069
	200	0.011	0.744	0.047	0.008	0.958	0.034
	250	0.010	0.764	0.036	0.003	0.979	0.019
323 K	150	0.006	0.841	0.026	0.004	0.980	0.016
	200	0.004	0.883	0.016	0.002	0.986	0.012
	250	0.004	0.903	0.012	0.281	0.047	0.672

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Table 7. The Activity Coefficients were Calculated by UNIFAC Model.

Solution	Temperature	Speed	γ_a^I	γ_b^I	γ_c^I	γ_a^{II}	γ_b^{II}	γ_o^{II}
Methanol	298	150	0.176	0.769	0.037	2.360	2.063	1.025
		200	0.032	0.794	0.004	2.398	2.098	1.020
		250	0.019	0.799	0.003	2.435	2.143	1.015
	313	150	0.149	0.789	0.006	2.143	2.655	1.016
		200	0.119	0.791	0.003	2.215	2.700	1.012
		250	0.112	0.792	0.003	2.191	2.685	1.013
	323	150	0.049	0.788	0.007	2.181	2.504	1.007
		200	0.028	0.788	0.001	2.222	2.515	1.006
		250	0.034	0.788	0.001	2.191	2.519	1.006
Acetic Acid	298	150	0.199	1.191	0.020	3.108	1.980	1.011
		200	0.132	1.002	0.001	3.150	1.996	1.009
		250	0.073	1.015	0.001	3.198	2.0161	1.006
	313	150	0.196	1.000	0.003	2.798	1.805	1.005
		200	0.152	0.999	0.013	2.857	1.824	1.002
		250	0.141	0.999	0.014	2.836	1.815	1.003
	323	150	0.055	0.999	0.037	2.705	1.701	1.006
		200	0.035	0.999	0.004	2.728	1.701	1.004
		250	0.042	0.999	0.006	2.719	1.711	0.999

17

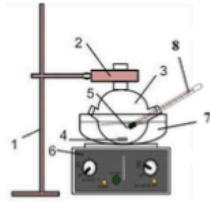


Figure 1. Extraction Equipment.

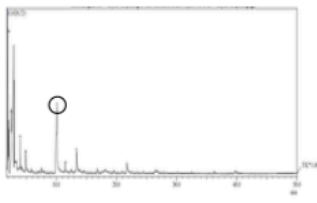


Figure 2. Analysis Result by GC-MS for the Bio-oil Produced by Coconut Shells Pyrolysis

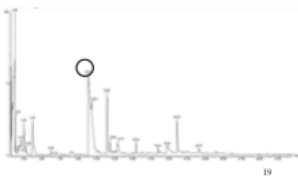


Figure 3. Analysis Result by GC-MS for the Bio-oil Produced by Coffee Shells Pyrolysis

Table 8. The Activity Coefficients were Calculated by NRTL Model

Solution	Temperature	Speed	γ_a^I	γ_b^I	γ_c^I	γ_a^{II}	γ_b^{II}	γ_o^{II}
Methanol	298	150	0.077	0.926	0.381	0.265	2.037	0.655
		200	0.064	0.482	0.337	0.001	0.039	0.065
		250	0.063	0.283	0.387	1.3×10^{-4}	0.001	7.6×10^{-4}
	313	150	4.47×10^{-4}	0.942	0.010	8.4×10^{-7}	1.050	0.044
		200	2.71×10^{-4}	0.986	0.056	1.5×10^{-11}	1.014	1.1×10^{-4}
		250	1.80×10^{-7}	0.944	0.039	4.9×10^{-14}	0.880	2.4×10^{-12}
	323	150	7.92×10^{-4}	0.980	0.010	3.6×10^{-4}	1.028	0.025
		200	3.48×10^{-4}	0.977	0.022	4.1×10^{-14}	1.006	4.1×10^{-12}
		250	9.44×10^{-4}	0.972	0.024	3.5×10^{-12}	0.937	1.1×10^{-9}
Acetic Acid	298	150	0.045	0.644	0.218	0.432	1.087	0.678
		200	0.039	0.331	0.210	0.015	0.361	0.552
		250	0.036	0.193	0.232	9.5×10^{-4}	0.017	0.027
	313	150	0.000	0.734	0.037	0.036	0.995	0.551
		200	0.003	0.732	0.050	0.030	0.999	0.444
		250	0.013	0.749	0.069	0.028	1.000	0.374
	323	150	0.000	0.734	0.037	0.035	1.139	0.011
		200	0.003	0.732	0.050	0.001	1.010	0.039
		250	0.013	0.749	0.069	8.9×10^{-4}	0.694	9.6×10^{-7}

Table 8. Sum of the Square of the Error (SSE)

Model	Temperature	SSE	
		Methanol	Acetic Acid
UNIFAC	298	0.087	0.132
	313	0.065	0.083
	323	0.053	0.084
NRTL	298	0.024	0.065
	313	0.026	0.544
	323	0.015	0.017

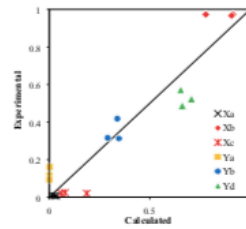


Figure 4. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 298 K.

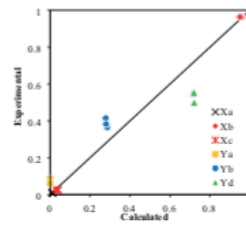


Figure 5. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 313 K.

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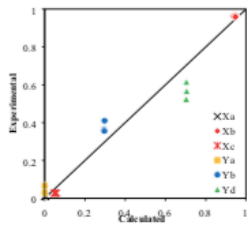


Figure 6. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 323 K.

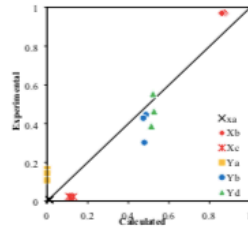


Figure 8. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 313 K.

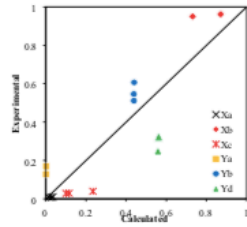


Figure 7. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 298 K.

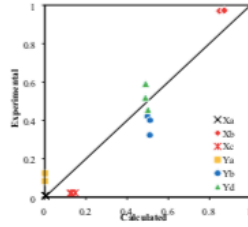


Figure 9. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 323 K.

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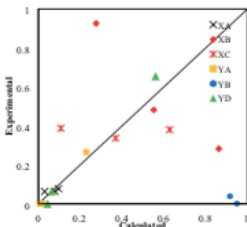


Figure 10. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 298 K.

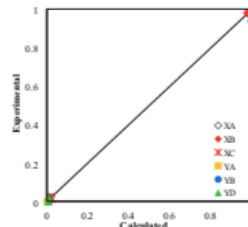


Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 323 K.

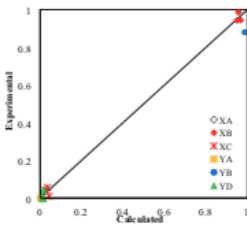


Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 313 K.

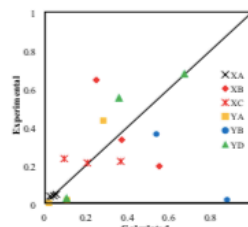


Figure 13. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 298 K.

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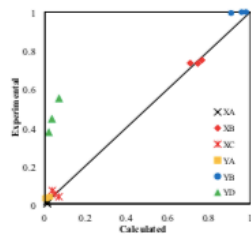


Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 313 K

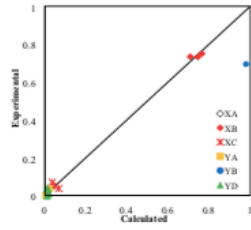


Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Acetic Acid by NRTL Model at 323 K

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Comments from the editors and reviewers:

-Reviewer 1

-

In this study, the author used the liquid-liquid extraction method to separate the phenol compounds from coconut shell pyrolysis bio-oils. The results show that the correlation between experimental data and calculations shows that the UNIFAC equilibrium model can be used to predict the liquid-liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the NRTL equilibrium model can be used to predict liquid-liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells. Generally, this study presents some important data on the development of liquid-liquid equilibrium in the extraction process towards pyrolysis bio-oil. This paper fits into the scope of liquid-liquid equilibrium in the extraction process. However, before that, the following minor points should be carefully responded and revised.

Specific comments:

1. Abstract

"34% phenol"

"The content of phenol compounds in corrosive bio-oils still quite high, **the use of this bio-oil** directly will cause various difficulties in the combustion system due to high viscosity"

Please give the full name for "UNIFAC equilibrium" and "NRTL equilibrium"

2. Introduction

Paragraph 1 Please add the related contents and references in this paragraph

3. 2.2 Equilibrium Procedure

"For the first stage, the aqua was used as a polar solvent, and chloroform as a non-polar solvent." may be better

What is "aquadest"? Error

"was conduct" should be "was conducted"

4. Please revise the x-coordinate and y-coordinate of Figure 7-11, and obtain the correct format

5. You mentioned "Gas Chromatography-Mass Spectrometry (GC-MS) analysis used to determine total content of phenol in coconut shell bio-oil." Please add the detection details.

-Reviewer 2

1. According to the meaning of context, "In Indonesia, in addition to the shell, one of the available biomass resources is coconut shell." should be replaced by "In Indonesia, in addition to the coffee shell, one of the available biomass resources is coconut shell." in the introduction.
2. In the section 2.2, why was the extraction conducted with ice-bath method at 4-5 °C? Is this operating temperature suitable for industrial applications?
3. In the section 3.1, "The results of GC-MS bio-oil analysis of coconut shell pyrolysis results can be seen in Figures 2 and 3." should be rewritten to understand better.
4. There were some mistakes in writing. Such as :
In Table 3 and Table 4, "Procentage" should be replaced by "Percentage".
In the section 3.2, "temperatur" should be rewritten as "temperature".

In the section 3.2, "temperatur" should be rewritten as "temperature".

5. In the section 3.1, there were two sentences, "It shows that coconut shell bio-oil contained total phenol for 40.01%." and "The total phenolic compounds contained in bio-oil is 47.03% (coconut shell)." . The percentages of total phenolic compounds pointed out in these sentences should be consistent.
6. In general, the speed of stirring has no effect on liquid-liquid equilibrium. The speed of stirring has a significant effect on the composition of liquid-liquid equilibrium from the Table 5 and Table 6. Why?
7. The data were too few to describe the liquid-liquid equilibrium well. Enough data are required.
8. In the section 3.3, "This is shown by the value of SSE (Sum of the Square of the Error) approaching the value of 0." This expression is inappropriate because the errors between calculated values and experimental data are not small from Fig.3 to Fig.15.
9. In the conclusion, the author pointed out that the UNIFAC modeling provides a good correlation to liquid-liquid equilibrium data under conditions of temperature of 323 K. Then, can the UNIFAC model be used to describe the liquid-liquid equilibrium under other temperatures? There is the same problem for the NRTL equation.

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Liquid Phase Equilibrium of Phenol Extraction from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models

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2. In the section 2.2, why was the extraction conducted with ice-bath method at 4-5 °C? Is this operating temperature suitable for industrial applications?

3. In the section 3.1, "The results of GC-MS bio-oil analysis of coconut shell pyrolysis results can be seen in Figures 2 and 3." should be rewritten to understand better.

4. There were some mistakes in writing. Such as :

In Table 3 and Table 4, "Percentage" should be replaced by "Percent".

In the section 3.2, "temperatur" should be rewritten as "temperature".

5. In the section 3.1, there were two sentences, "It shows that coconut shell bio-oil contained total phenol for 40.01%," and "The total phenolic compounds contained in bio-oil is 47.03% (coconut shell).". The percentages of total phenolic compounds pointed out in these sentences should be consistent.



6. In general, the speed of stirring has no effect on liquid-liquid equilibrium. The speed of stirring has a significant effect on the composition of liquid-liquid equilibrium from the Table 5 and Table 6. Why?





7. The data were too few to describe the liquid-liquid equilibrium well. Enough data are required.

8. In the section 3.3, "This is shown by the value of SSE (Sum of the Square of the Error) approaching the value of 0." This expression is inappropriate because the errors between calculated values and experimental data are not small from Fig.3 to Fig.15.

9. In the conclusion, the author pointed out that the UNIFAC modeling provides a good correlation to liquid-liquid equilibrium data under conditions of temperature of 323 K. Then, can the UNIFAC model be used to describe the liquid-liquid equilibrium under other temperatures? There is the same problem for the NRTL equation.

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Manuscript Details

Manuscript number	CJCHE_2018_233_R1
Title	Liquid Phase Equilibrium of Phenol Extraction from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models
Short title	Liquid Phase Equilibrium of Phenol Extraction from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models
Article type	Research Paper

Abstract

Utilization of biomass as a new and renewable energy source is being actively conducted by various parties. One of the technology for utilizing or converting biomass as an energy source is pyrolysis, to convert biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from vapors phase of biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. The total phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). The content of phenol compounds in corrosive bio-oils still quite high, the use of this bio-oil directly will cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive, and unstable. Phenol compounds have some benefits as one of the compound for floor cleaner and disinfectant which contained in bio-oil. The correlation between experimental data and calculations shows that the UNIQUAC Functional-group Activity Coefficients (UNIFAC) equilibrium model can be used to predict the liquid-liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the Non Random Two Liquid (NRTL) equilibrium model can be used to predict liquid-liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells.

Keywords	biomass, pyrolysis, bio-oil, UNIFAC, NRTL
Manuscript category	Chemical engineering thermodynamics
Corresponding Author	dewi selvia fardhyanti
Order of Authors	dewi selvia fardhyanti, bayu triwibowo, Heri Istanto, Muhammad Khuzni Anajib, Amalia Larasati, Windy Oktaviani
Suggested reviewers	Catherine Peters, dhori hartanto, Ming-Jer Lee

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PART 1:

Journal Name:	Chinese Journal of Chemical Engineering
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Title of the Manuscript:	Liquid Phase Equilibrium of Phenol Extraction from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models

PART 2:

Reviewer A (Yellow Highlight)	Authors' response to reviewer's comments
<p>1. Abstract</p> <p>"34% phenol"</p> <p>"The content of phenol compounds in corrosive bio-oils still quite high, the use of this bio-oil directly will cause various difficulties in the combustion system due to high viscosity"</p> <p>Please give the full name for "UNIFAC equilibrium" and "NRTL equilibrium"</p>	<p>1. The manuscript has been revised.</p> <p>Line 19.</p> <p>Line 19-21.</p> <p>Line 24-25 and 26-27.</p>
<p>2. Introduction</p> <p>Paragraph 1 please add the related contents and references in this paragraph</p>	<p>2. The manuscript has been revised.</p>
<p>3. 2.2. Equilibrium Procedure</p>	<p>3. The manuscript has been revised.</p>

<p>"For the first stage, the aqua was used as a polar solvent, and dibutylform as a non-polar solvent." may be better</p> <p>What is "aquadet"? Error</p> <p>"was conduct" should be "was conducted"</p>	<p>Line 128-129.</p> <p>Line 129.</p>
<p>4. Please revise the x-coordinate and y-coordinate & x/y/z; Figure 7-11&A/F/D9, and obtain the correct format.</p>	<p>4. The manuscript has been revised.</p> <p>Line 440-522.</p>
<p>5. You mentioned "Gas Chromatography-Mass Spectrometry (GC-MS) analysis used to determine total content of phenol in coconut shell bio-oil". Please add the detection details.</p>	<p>5. The manuscript has been revised.</p> <p>Line 145-155.</p>
<p>PART 3:</p> <p>Reviewer B (Green Highlight)</p>	<p>Authors' response to reviewer's comments</p>
<p>1. According to the meaning of context, "In Indonesia, in addition to the shell, one of the available biomass resources is coconut shell." should be replaced by "In Indonesia, in addition to the coffee shell, one of the available biomass resources is coconut shell." in the introduction.</p>	<p>1. The manuscript has been revised.</p> <p>Line 56.</p>

<p>2. In the section 2.2, why was the extraction conducted with ice-bath method at 4-5°C? Is this operating temperature suitable for industrial applications?</p>	<p>2. This operating temperature has not been yet used for industrial application. It is still used for laboratory system.</p>
<p>3. In the section 3.1, "The results of GC-MS bio-oil analysis of coconut shell pyrolysis results can be seen in Figures 2 and 3" should be rewritten to understand better.</p>	<p>3. The manuscript has been revised.</p> <p>Line 145-155.</p>
<p>4. There were some mistakes in writing. Such as: In Table 3 and Table 4, "Percentage" should be replaced by "Percentage". In the section 3.2, "temperature" should be rewritten as "temperature".</p>	<p>4. The manuscript has been revised.</p> <p>Line 152.</p> <p>Line 161 and 163.</p>
<p>5. In the section 3.1, there were two sentences, "It shows that coconut shell bio-oil contained total phenol for 40.01%" and "The total phenolic compounds contained in bio-oil is 47.03% (coconut shell)". The percentages of total phenolic compounds pointed out in these sentences should be consistent.</p>	<p>5. The manuscript has been revised.</p> <p>Line 155-156.</p>

<p>6. In general, the speed of stirring has no effect on liquid-liquid equilibrium. The speed of stirring has a significant effect on the composition of liquid-liquid equilibrium from the Table 5 and Table 6. Why?</p>	<p>6. In general, the speed of stirring has no effect for LLE. It was showed from the Table 5 and Table 6 that almost all the compositions of the extract phase and raffinate phase for each temperature has approaching the same value. For example: T=298 K, X₁ for speed 150 is 0.007, for speed 200 is 0.007, and for speed 250 is 0.009, etc. Only a few data showed a big difference.</p>	<p>8. In the section 3.3, "This is shown by the value of SSE (Sum of the Square of the Error) approaching the value of 0." This expression is inappropriate because the errors between calculated values and experimental data are not small from Fig.3 to Fig.15.</p>	<p>8. The manuscript has been revised. Line 310-311. Table 8 showed that the value of SSE is 0.015 - 0.554.</p>
<p>7. The data were too few to describe the liquid-liquid equilibrium well. Enough data are required.</p>	<p>7. One of the important thing to describe the LLE is the Activity Coefficient. It was showed the ideal conditions for each component in the system, UNIFAC and NRTL is used to calculate the activity coefficient for each component and calculated the calculated data, and then the calculated data and experimental data are compared to describe and showed the better conditions for each temperature. The activity coefficients for each model are shown in Table 7 and Table 8. Line 267-277 showed the explanation of the activity coefficient of each model and conditions.</p>	<p>9. In the conclusion, the author pointed out that the UNIFAC modeling provides a good correlation to liquid-liquid equilibrium data under conditions of temperature of 323 K. Then, can the UNIFAC model be used to describe the liquid-liquid equilibrium under other temperatures? There is the same problem for the NRTL equation.</p>	<p>9. The manuscript has been revised. Line 326-331. It was explained in line 314-323. So, the UNIFAC and NRTL model could not be used to describe the liquid-liquid equilibrium under or up 323 K.</p>

1 **Liquid Phase Equilibrium of Phenol Extraction**
2 **from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models**
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11
12 **ABSTRACT**
13
14 Utilization of biomass as a new and renewable energy source is being actively conducted by various parties.
15 One of the technology for utilizing or converting biomass as an energy source is pyrolysis, to convert
16 biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from vapors phase of
17 biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of
18 hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. The total
19 phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). The content of
20 phenol compounds in corrosive bio-oils still quite high, the use of this bio-oil directly will cause various
21 difficulties in the combustion system due to high viscosity, low calorific value, corrosive, and unstable.
22 Phenol compounds have some benefits as one of the compound for floor cleaner and disinfectant which
23 contained in bio-oil.
24 The correlation between experimental data and calculations shows that the UNIQUAC Functional-group
25 Activity Coefficients (UNIFAC) equilibrium model can be used to predict the liquid-liquid equilibrium in
26 the phenol extraction process of the coconut shell pyrolysis bio-oil. While the Non Random Two Liquid
27 (NRTL) equilibrium model can be used to predict liquid-liquid equilibrium in the extraction process of
28 phenol from bio-oil pyrolysis of coffee shells.
29
30
31 **Keywords:** biomass, pyrolysis, bio-oil, UNIFAC, NRTL.

65 conversion process [11]. Chemical content of coconut shell is cellulose (34%), hemicellulose (21%),
66 lignin (27%) and ash (18%) [12].
67
68 Before converted into bio-oil, coconut shell waste and coffee shell need to be treated first. It takes a
69 process before converting the waste of coffee shell into bio-oil which is pyrolysis. Pyrolysis is the
70 process of decomposition of a substance or material that is done at a relatively high temperature.
71 Pyrolysis results in the form of gas, liquid and solid. The solid result of this process is char (char). Gas
72 and liquid products can be used as a source of bio-oil. Bio-oil can be used as an alternative fuel that is
73 easier to store and distribute than biomass in solid form. Bio-oil made from coconut shell material using
74 slow pyrolysis with pyrolysis temperature 250-300 °C produces liquid which very acidic and corrosive.
75 The largest organic component in bio-oil is the lignin derivative of phenol, alcohol, organic acids and
76 carbonyl compounds such as ketones, aldehydes and esters [13]. The nature of the acid is caused by the
77 content of the dominant phenol compounds.
78
79 Phenol compounds in addition to causing the smell of smoke in bio-oil, also showed antioxidant
80 activity and corrosive [14]. Therefore, the use of this bio-oil will directly cause various difficulties in
81 the combustion system due to high viscosity, low calorific value, corrosive and unstable [6].
82
83 Phenol needs to be extracted from bio-oil to reduce its corrosive properties. In addition, phenol is a
84 compound that has a high economic value because of its expensive and widely used. Phenols have many
85 important uses such as disinfectants, resins, pesticides, explosives, and drugs. The method that can be
86 used to extract phenol compounds from bio-oil is by liquid-liquid extraction method using methanol
87 solvent. This has been done by Jazbinsk [15] on the isolation of phenol compounds from bio-oils from
88 pyrolysis of forest waste and Mantilla [16] on the extraction of phenol compounds from bio-oils from
89 pyrolysis of agricultural waste. Dahlia [17] conducted research on liquid-liquid extraction of phenol
90 compound from bio-oil of pyrolysis resulted of palm oil empty bunch using methanol solvent. The yield
91 of phenol resulting from this extraction is 40%. In this study, it is expected that the liquid-liquid
92 extraction method can be used to separate the phenol compounds from coconut shell pyrolysis bio-oils.
93 The purpose of phenol extraction is to improve the quality of bio-oil and reduce the corrosivity, so when
94 used as fuel, it will cause no damage to the engine.
95
96 Biomass pyrolysis converted to bio-oil studies have been conducted. Bio-oil produced from the
97 pyrolysis process can be used as fuel for engines, turbines and boilers. The bio-oil which processed
98 further by liquid-liquid extraction process will reduce the corrosive nature of bio-oil and obtained
99 chemicals which one of them is phenol. Research on extraction of phenol compounds from bio-oil is
100 still very limited. In addition, thermodynamic modelling study on liquid-liquid equilibrium of phenol
101 extraction process from bio-oil still needs to be investigated further.

32 **1. INTRODUCTION**
33

34 The role of fossil fuels still be crucial, but their effects will gradually be over taken by new and
35 renewable energy sources. Along with the depletion of world oil reserves and environmental issues,
36 various researches on renewable energy are being actively conducted by various parties. Therefore,
37 technology needs be able to overcome the problem. One technology that can be used is pyrolysis
38 technology where the material used comes from organic waste (biomass). Pyrolysis technology can
39 overcome organic waste problems and make the environment healthy also convert it into value-added
40 end product.

41 Biomass is an organic material derived from plants, algae, animal waste, industrial waste, forestry, and
42 agricultural waste [1]. Biomass can be used as pyrolysis raw material to produce liquid (bio-oil), gas and
43 solid products. Various kinds of biomass waste can be used as raw materials for the manufacture of bio-
44 oils, such as corncobs [2], sawdust and wheat bran [3], castor [4], oak [5], durian shell [6] and coconut
45 shell [7].

46 Indonesia is the third largest coffee producer in the world after Brazil and Vietnam by contributing
47 about 6% of the world's total coffee production, also world's fourth largest coffee exporter with a market
48 share of around 11% across the world. Coffee is one of the plantations with high economic value besides
49 chocolate and tea. Based on data from Indonesian Ministry of Forestry and Plantation, Indonesia's coffee
50 production in 2013 reached 691,160 tons with a plantation area of 47,800 Ha [8]. Waste from coffee
51 processing that abundant and not utilized optimally is the coffee shell. Large waste of coffee shells
52 derived from plantations was approximately 11.08% which potentially cause environment hazard.
53 Coffee shells are a waste that has high lignocellulose content which have potential to be used as bio-oil.
54 Bio-oil is a blackish liquid fuel derived from biomass such as wood, coffee shells and other biomass.
55 Bio-oil contains lignin components (23.4%), cellulose (34.2%) and hemicellulose (24.5%) [9].

56 In Indonesia, in addition to the coffee shell, one of the available biomass resources is coconut shell
57 Availability of coconut shell can be seen from the production of coconut fruit in Indonesia. Coconut is
58 one of Indonesia's main commodities with an average production of 3,000,000 tons / year [10]. The
59 utilization of coconut is very diverse, but for the utilization and way of handling waste is still limited.
60 Coconut fruit can be used as food in daily life for the purpose of consumed directly. Coconut shells
61 nowadays, lots of it become garbage that can not be utilized. One of the waste handling is to utilize
62 coconut shell waste as a base material in the manufacture of bio-oil. Characterization studies on coconut
63 shells have been done through proximate, ultimate, and thermo gravimetric (TG) analyzes. The results
64 of this analysis indicate that coconut shell has a high potential to produce fuel fluid by pyrolysis

98 To design extraction columns required various information such as liquid-liquid equilibrium data
99 (Liquid-liquid Equilibrium). Thermodynamic models such as Equation of State or Activity Coefficient
100 are used to correlate the experimental data to obtain optimal interaction parameters. Optimal parameters
101 can be used to predict liquid-liquid equilibrium data to produce an equilibrium graph that is used to
102 design extraction columns [18]. The study on liquid-liquid equilibrium include NRTL, e-NRTL,
103 UNIQUAC, and UNIFAC.

104 Given the importance of thermodynamic modeling in this liquid-liquid extraction process and the price
105 and benefits of phenol in the industrial world, this research becomes an important thing to do. From the
106 equilibrium data, the optimum condition will be obtained where the highest yield of extract obtained in
107 the extraction process of phenol compound from bio-oil from coconut pyrolysis. Liquid-liquid
108 equilibrium data of the components of the system can be used in the design of the equipment in
109 separation process.

110 **2. EXPERIMENTAL SECTION**

111 **2.1 Chemicals**

112 All reagents used here are A.R. grade and used as received without further purification. Methanol,
113 acetic acid, and chloroform were purchased from e-Merck (Germany). The distilled water is available at
114 the laboratory. Coffee shell and coconut shell were obtained from public market in Semarang.

115 **2.2 Equilibrium Procedure**

- 116 Instruments used in this study:
117 a) Shaker bath, product by Memmert WB14, SV1422, Schwabach, Germany.
118 b) Gas Chromatography (GC), product by GC 6820 Agilent Technologies, column: Rastek RXi-
119 5MS002E.
120 c) Gas Chromatography-Mass Spectroscopy (GC-MS), product by GCMS-QP2010S Shimadzu,
121 column: Rastek RXi-5MS.

122 The material systems and experimental conditions for the equilibrium extraction are summarized in
123 Table 1.

124 **Table 1. Material Systems and Conditions for Equilibrium Extraction**

125 The bio-oil feed solution that containing phenol was extracted using two-stage extraction. The first
126 stage uses distilled water as a polar solvent and chloroform as a non-polar solvent. Extraction was
127 conducted with ice-bath method at 4-5°C, 200 rpm and stirring time for 2 hours. The extracts obtained at
128 this first stage are used as feeds on the second stage extraction. At the second stage extraction, each
129 extract was extracted using a methanol solvent under operating conditions shown in Table 1.

134
135 **2.3 Analysis and Characterization**
136
137 After equilibrium attained (60 minutes), the mixtures were poured into a separating funnel, settled for an
138 hour and separated into two phases. Then the two phases were weighed. The extract phase and the
139 raffinate phase of each extraction processes are analyzed by GC to obtain the concentration of each
140 component. The principal conditions of this analysis are shown in Table 2.

141 **Table 2. Conditions of Analysis Using GC**

142
143 **3. RESULTS AND DISCUSSION**
144

145 **3.1 Bio-Oil Content**

146 The chromatograms of the GC-MS analysis are shown in Fig. 2 and 3. The results of the GC-MS
147 analysis of the bio-oil showed that they contained more than six chemical compounds such as ethyl ester,
148 phenol, furfural, acetic acid, and others.

149 **Figure 2. The Chromatogram of Bio-Oil Produced from Coconut Shell Pyrolysis**

150 **Table 3. The Components of Bio-oil Produced from Coconut Shell Pyrolysis**

151 **Figure 3. The Chromatogram of Bio-Oil Produced from Coffee Shell Pyrolysis**

152 **Table 4. Component of Bio-oil Produced from Coffee Shell Pyrolysis**

153 Table 3 and 4 represent that bio-oil contains more than 6 main components. The percentage (%) of the
154 composition of the bio-oil analysis by GC-MS was obtained from the percentage area of the peak or the
155 height of the peak in the chromatograms. Using a method of GC-MS, this work presents a composition
156 of the bio-oil generated in the experiment. The total phenolic compounds contained in bio-oil is 47.03%
157 (coconut shell) and 45% (coffee shell). It was caused by the pyrolysis processing at 300-350°C (low
158 temperature carbonization) that produce more substances contain ester and phenolic compounds. The
159 composition of biomass such as hemicellulose, cellulose and lignin will be oxidized to phenol as the main
160 component of the bio-oil.

161 **3.2 Equilibrium data on phenol extraction of bio-oil from coconut pyrolysis results**

162 In this research the variables studied were temperature, speed of stirring and type of solvents (aqueous
163 methanol and aqueous acetic acid). Generally, all extracts are rich of phenol but the mole fraction
164 significantly depends on extraction conditions: temperature, speed of stirring and type of solvents.
165 Methanol and acetic acid need to be mixed with water because their polarities are not strong enough to
166 form immiscible mixture with bio-oil [19].

166 The mole fractions of any components in the raffinate phase and the extract phase are constant after 60
167 minutes, so the system is assumed to be in equilibrium. The amount of water transferred from the solvent
168 into the raffinate phase was negligible [19].

169 In this work, we are reported experimental data for the phenol extraction from bio-oil produced from
170 coconut shell and the phenol extraction from bio-oil produced from coffee shell.

171 Table 5 and 6 show the equilibrium of multi-components extraction data for aqueous acetone and
172 aqueous methanol solvent in the extract phase and raffinate phase.

173 **Table 5. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous
174 Methanol from Bio-oil Produced from Coconut Shell.**

175 **Table 6. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Acetic
176 Acid from Bio-oil Produced from Coffee Shell.**

177 **3.3 Liquid-liquid Equilibrium for The Phenol Extraction Process using UNIFAC dan NRTL
178 Models.**

179 The UNIFAC and NRTL models were used to correlate the experimental data obtained for the systems
180 under study. UNIFAC equation and NRTL equation (Smith, 1996) were used to correlate experimental phase
181 equilibrium data. The activity coefficient models applied in this study are as follows:

182 a) UNIFAC: [20]

183 UNIFAC is a UNIQUAC Functional-group Activity Coefficients model based on the UNIQUAC
184 equation by using functional groups of a molecule contained in the mixture to calculate the activity
185 coefficients. This model was developed by Fredenslund, Jones, and Prausnitz (1975). The UNIFAC
186 modeling is a continuation of the UNIQUAC modeling development. This model has the advantage of
187 performing phase equilibrium estimation even in the absence of experimental data. This thermodynamic
188 model is used to predict the estimation of non-electrolyte activity in non-ideal mixtures. The UNIFAC
189 model calculates the activity coefficients by using functional groups present in the molecules that form
190 the liquid mixture. If the value of the component activity coefficient has a value close to one, this indicates
191 that the component is in the ideal state in the system [21]. The correlation in the UNIFAC model is used
192 in order to reduce the number of binary interactions measured to predict the state of the system. The
193 correlation for the UNIFAC model is shown as follows:

194
$$\ln \gamma_i = \ln \gamma_i^f + \ln \gamma_i^R \quad (1)$$

195
$$\ln \gamma_i^f = 1 - J_i + \ln J_i - 5q_i \left(1 - \frac{1}{L_i} + \ln \frac{1}{L_i} \right) \quad (2)$$

196
$$\ln \gamma_i^R = q_i \left[1 - \sum_k \left(\theta_k \frac{r_k}{r_i} - e_{ik} \ln \frac{r_k}{r_i} \right) \right] \quad (3)$$

197
$$J_i = \frac{r_i^2 p_i}{\sum_j r_j^2 p_j} \quad (4)$$

198
$$L_i = \frac{r_i^2 q_i}{\sum_k r_k^2 q_k} \quad (5)$$

199
$$r_i = \sum_k v_k^{(i)} R_k \quad (6)$$

200
$$q_i = \sum_k v_k^{(i)} Q_k \quad (7)$$

201
$$e_{ik} = \frac{v_i^{(i)} Q_k}{q_i} \quad (8)$$

202
$$\beta_{ik} = \sum_m e_{im} \tau_{mk} \quad (9)$$

203
$$\theta_k = \frac{\sum_i r_i \beta_{ik}}{\sum_i r_i} \quad (10)$$

204
$$s_k = \sum_m \beta_{mk} \tau_{mk} \quad (11)$$

205
$$\tau_{mk} = \exp \left(-\frac{u_{mk}}{T} \right) \quad (12)$$

206 Where:

207 $\ln \gamma_i^C$ = Combinatorial component

208 $\ln \gamma_i^R$ = residual component

209 J_i = component surface area component (i)

210 L_i = component volume fraction (i)

211 r_i = pure component surface area parameter (i)

212 x_i = pure component mol fraction (i)

213 q_i = pure component volume parameter (i)

214 $v_k^{(i)}$ = number of sub-group (k) on component (i)

215 R_k = parameter area for sub-group (k) ($R_k = A_{sk} / (2.5 \times 10^3)$, where A_{sk} is number of sub-group
216 van der Waals surface area)

217 Q_k = number of parameter volume sub-group (k) ($Q_k = V_{sk} / 15.17$, where V_{sk} is number of Van
218 der Waals volume sub-group)

219 a_{mk} = group of interaction parameter from group m and k [18]

220 s_i = liquid mol fraction of component (i)

221 UNIFAC equation can be obtained to :

222
$$\ln \gamma_i = \ln \left[1 - J_i + \ln J_i + 5q_i \left(1 - \frac{1}{L_i} + \ln \frac{1}{L_i} \right) \right] + \ln \left[1 - \sum_k \left(\frac{\sum_i r_i \beta_{ik}}{\sum_i r_i} \frac{r_k}{r_i} - e_{ik} \ln \frac{r_k}{r_i} \right) \right] \quad (13)$$

223
224
225 b) NRTL: [20]

226 Non Random Two Liquid (NRTL) is derived from the Scott two liquid model and it is assumed that
227 randomness is the same as the model used on the Wilson model. The NRTL model has a non-random
228 parameter (α_{12}) making it possible to use in a wide variety of mixtures. The NRTL model uses only
229 binary parameters to calculate the phase equilibrium properties.

230 The NRTL (Non Random Two-Liquid) equation was first developed by Renon and Prausnitz (1968).
231 The NRTL model is derived from the Scott two liquid model and it is assumed that the non-random
232 parameter is the same as the model used in the Wilson Model. The NRTL equation is developed based
233 on a two-fluid theory with the addition of a non-random parameter (α), which makes the equation
234 applicable to different types of liquid-liquid mixture and equilibrium by choosing the appropriate price.
235 The parameter estimation for this equation is done by setting the α value at a certain value and
236 subsequently optimizing the objective function to obtain two optimal parameter interactions. The
237 recommended α value is 0.2 - 0.47. The value of α is often adjusted to certain conditions, but usually the
238 value of $\alpha = 0.3$. Unlike the Wilson equation which contains two adjustable parameters, the NRTL
239 equation contains three parameters, so the NRTL equation can be an alternative model if other models
240 are inadequate. This equation can also estimate the equilibrium of multicomponent systems without
241 additional parameters.

242 The NRTL equation can represent liquid-liquid equilibrium for both binary and multicomponent
243 systems only with binary parameters. The equation is simpler than the UNIQUAC equation, but it has
244 disadvantage because it contains three parameters for each pair of components. The third parameter (α)
245 is estimated based on the state of the component and sometimes for certain substances the extreme
246 number is obtained. The NRTL equation is expressed:

247
$$\frac{G^E}{RT} = \sum_i x_i^2 \frac{\sum_k \sum_l v_k v_l \tau_{kl}}{\sum_k \sum_l v_k v_l} \quad (14)$$

248
$$\tau_{ij} = \frac{g_{ij} - g_{ji}}{RT} \quad (15)$$

249
$$g_{ii} = g_{jj} = g_{kk} = 0 \quad (16)$$

250
$$G_{ij} = \exp(-\alpha_{ij} \tau_{ij}) \quad (17)$$

251
$$\alpha_{ij} = \alpha_{ji} \quad (18)$$

252
$$\ln \gamma_i = \frac{\sum_k \sum_l v_k v_l G_{kl}}{\sum_k \sum_l v_k v_l} + \sum_j \frac{x_j}{x_i} \frac{g_{ij}}{\sum_k \sum_l v_k v_l} \tau_{ij} - \sum_k \frac{v_k g_{ki}}{\sum_k \sum_l v_k v_l} \quad (19)$$

253
$$x_i = \exp \left(\frac{\sum_k \sum_l v_k v_l G_{kl}}{\sum_k \sum_l v_k v_l} + \sum_j \frac{x_j}{x_i} \frac{g_{ij}}{\sum_k \sum_l v_k v_l} \tau_{ij} - \sum_k \frac{v_k g_{ki}}{\sum_k \sum_l v_k v_l} \right) \quad (20)$$

$$254 \quad V_i = \exp \left(\frac{\sum_{j=1}^n \frac{V_i^L V_j^L}{V_i^L + V_j^L} + \sum_{j=1}^n \frac{V_i^L}{V_i^L + V_j^L} (v_j - v_i) - \frac{V_i^L V_j^L}{V_i^L + V_j^L} \right) \quad (21)$$

255 Where:

- 256 G^E = molar Gibbs energy
- 257 R = gas constant
- 258 T = temperature
- 259 x = liquid phase mol fraction
- 260 i, j, k = component of 1, 2, 3, ..., n
- 261 α = non-random parameter for NRTL
- 262 β, G = component parameter
- 263 γ = activity coefficient
- 264 τ = empirical constant for NRTL

265 **Table 7. The Activity Coefficients were Calculated by UNIFAC Model**

266 **Table 8. The Activity Coefficients were Calculated by NRTL Model**

267 Table 7 and 8 shows the value of activity coefficient on phenol extraction using methanol solvent
 268 obtained from calculation with UNIFAC and NRTL modeling, from coefficient value of activity on each
 269 component shows that methanol and acetic acid component in extract phase is in ideal condition because
 270 the value of activity coefficient obtained close to one. The activity coefficient value of phenol and water
 271 activity in the extract phase is not close to one, it means that the phenol and water components are in less
 272 than ideal conditions in the system. The ideal conditions of the components in the system show that the
 273 component is dominant, so the properties of the mixture are closer to the dominant component properties
 274 either in the extract or raffinate phase. In raffinate it is known that the solvent has a coefficient of activity
 275 close to the one, indicating that the solvent is in ideal conditions in the system, while the phenol and bio-
 276 oil components are in less than ideal conditions. So non-ideality studies are very important for those
 277 systems [21].

278 **Figure 4. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 279 **using Aqueous Methanol by UNIFAC Model at 298 K.**

280 Where:

- 281 X_a : mole fraction of component in extract phase
- 282 X_b : mole fraction of solvent in extract phase
- 283 X_c : mole fraction of water in extract phase
- 284 Y_a : mole fraction of component in raffinate phase

10

315 the smallest SSE value compared to other conditions with SSE value of 0.053 (for UNIFAC model) and
 316 0.015 (for NRTL model). The smaller the SSE values obtained show that the correlation between the
 317 experimental data and the calculation data indicates a better correlation. Good correlation results show
 318 that equilibrium modeling of UNIFAC and NRTL can be used to predict liquid-liquid equilibrium in
 319 phenol extraction process from coconut shell pyrolysis bio-oil and coffee shell using methanol solvent
 320 and acetic acid at 323 K.

321 Figure 4-12 shows that the visual comparison of calculated and experimental mole fractions in the
 322 extract phase and the raffinate phase for the methanol solvent system by UNIFAC and NRTL at 323 K
 323 are better than the other conditions.

324
 325 **4. CONCLUSION**

326 The UNIFAC modeling provides a good correlation to liquid-liquid equilibrium data for ternary systems
 327 on phenol extraction of coconut shell pyrolysis bio-oil using methanol solvent under conditions of
 328 temperature of 323 K given by SSE of 0.053.

329 NRTL modeling gives a good correlation to liquid-liquid equilibrium data for ternary system on phenol
 330 extraction of bio-oil from pyrolysis of coffee shell using methanol solvent at temperature condition of
 331 323 K given by SSE of 0.015.

332 The activity coefficients of methanol, acetic acid, and phenol in the extract phase and raffinate phase
 333 are far from unity. It means that they are far from ideal condition. The activity coefficients of water and
 334 kerosene in their phases are close to one. It means that they are nearly ideal in their phases.

335
 336
 337 **ACKNOWLEDGMENTS**

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 340 Negeri Semarang, Nomor: 084/SP2/H/LT.DRPM/IV/2017.

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- 285 Y_b : mole fraction of solvent in raffinate phase
- 286 Y_d : mole fraction of bio-oil in raffinate phase

287 **Figure 5. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 288 **using Aqueous Methanol by UNIFAC Model at 313 K.**

289 **Figure 6. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 290 **using Aqueous Methanol by UNIFAC Model at 323 K.**

291 **Figure 7. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 292 **using Aqueous Acetic Acid by UNIFAC Model at 298 K.**

293 **Figure 8. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 294 **using Aqueous Acetic Acid by UNIFAC Model at 313 K.**

295 **Figure 9. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 296 **using Aqueous Acetic Acid by UNIFAC Model at 323 K.**

297 **Figure 10. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 298 **using Aqueous Methanol by NRTL Model at 298 K.**

299 **Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 300 **using Aqueous Methanol by NRTL Model at 313 K.**

301 **Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 302 **using Aqueous Methanol by NRTL Model at 323 K.**

303 **Figure 13. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 304 **using Aqueous Acetic Acid by NRTL Model at 298 K.**

305 **Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 306 **using Aqueous Acetic Acid by NRTL Model at 313 K.**

307 **Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction**
 308 **using Acetic Acid by NRTL Model at 323 K.**

309 The result of correlation of liquid-liquid equilibrium data by using UNIFAC and NRTL modeling gives
 310 good agreement between experimental data and calculation. This is shown by the value of SSE (Sum of
 311 the Square of the Error) is very small. The SSE values in UNIFAC and NRTL modeling can be seen in
 312 Table 8. The value of SSE is 0.015 - 0.554.

313 **Table 8. Sum of the Square of the Error (SSE)**

314 **Based on Table 8, it can be seen that extraction with methanol solvent with temperature of 323 K has**

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 387 components extraction by thermodynamic models, *Int. J. Thermodyn.* 19(2) (2015) 110-119.

388 **Table 1. Material Systems and Conditions for Equilibrium Extraction.**

	Quantity
feed	bio-oil
feed volume	2.5 mL
solvent	aqueous methanol
	aqueous acetic acid
mole fraction of water in solvent	0.2
solvent-feed mass ratio	1
shaking time	60 minutes
amplitude of shaking	150, 200, 250 rpm
temperature	298, 313, 323 K.

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 391
 392 **Table 2. Conditions of Analysis Using GC**
 393

Column: Rastök RXG-SMS		
Column:		
Inner diameter	[m]	3.2 x 10 ⁻⁴
Length	[m]	30
Carrier gas		He
Split ratio	[-]	153
Flow rate	[cm ³ /sec]	26.6
Sample volume	m ³	1.10 ⁻⁹
Injection temperature	[K]	553
Column temperature	[K]	313 - 573
Pressure column	kPa	10.0
Column flow	mL/min	0.54
Detector (FID) temperature	[K]	573

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402 **Table 3. The Main Components of Bio-oil Produced by Pyrolysis of Coconut Shells**

Peak	Component	Percentage
1	Ethyl ester	37.60
2	Formic acid	4.02
3	1-Hydroxy-2-butanone	3.89
4	Furfural	5.45
5	Phenol	40.01
7	2-Methoxy-phenol	7.02

404
 405
 406
 407
 408 **Table 4. The Main Components of Bio-oil Produced by Pyrolysis of Coffee Shells**

Peak	Component	Percentage
1	Urea	5%
2	Acetic acid	26%
5	Furfural	6%
6	Phenol	34%
7	Phenol, 2-methoxy-	7%
8	Phenol, 2,6-dimethoxy	4%

409
 410
 411
 412
 413 **Table 5. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Methanol from Bio-oil Produced from Coconut Shell**

Temperature	Speed	X _A	X _B	X _C	Y _A	Y _B	Y _D
298 K	150	0.007	0.973	0.020	0.161	0.316	0.524
	200	0.007	0.973	0.019	0.116	0.313	0.572
	250	0.009	0.967	0.024	0.094	0.488	0.418
313 K	150	0.009	0.963	0.027	0.085	0.549	0.365
	200	0.008	0.970	0.022	0.086	0.414	0.501
323 K	250	0.010	0.963	0.027	0.062	0.555	0.383
	150	0.010	0.962	0.028	0.069	0.567	0.363
323 K	200	0.009	0.965	0.026	0.065	0.525	0.410
	250	0.012	0.959	0.029	0.029	0.615	0.356

414
 415
 416 **Table 6. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Acetic Acid from**

417 **Bio-oil Produced from Coffee Shell**

Temperature	Speed	X _A	X _B	X _C	Y _A	Y _B	Y _D
298 K	150	0.054	0.247	0.365	0.281	0.047	0.672
	200	0.042	0.370	0.204	0.107	0.537	0.357
	250	0.024	0.551	0.093	0.019	0.878	0.104
313 K	150	0.012	0.707	0.068	0.025	0.907	0.069
	200	0.011	0.744	0.047	0.008	0.958	0.034
323 K	250	0.010	0.764	0.036	0.003	0.979	0.019
	150	0.006	0.841	0.026	0.004	0.980	0.016
323 K	200	0.004	0.883	0.016	0.002	0.986	0.012
	250	0.004	0.903	0.012	0.001	0.947	0.007

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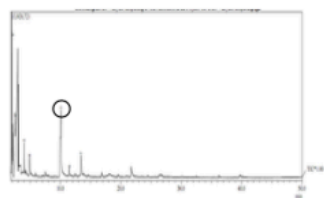
421 **Table 7. The Activity Coefficients were Calculated by UNIFAC Model.**
422

Solution	Temperature	Speed	γ_a^I	γ_a^II	γ_c^I	γ_a^{II}	γ_a^{III}	γ_a^{IV}
Methanol	298	150	0.176	0.769	0.037	2.360	2.063	1.025
		200	0.032	0.794	0.004	2.398	2.098	1.020
		250	0.019	0.799	0.003	2.435	2.143	1.015
	313	150	0.149	0.789	0.006	2.143	2.655	1.016
		200	0.119	0.791	0.003	2.215	2.700	1.012
		250	0.112	0.792	0.003	2.191	2.685	1.013
	323	150	0.049	0.788	0.007	2.181	2.504	1.007
		200	0.028	0.788	0.001	2.222	2.515	1.006
		250	0.034	0.788	0.001	2.191	2.519	1.006
Acetic Acid	298	150	0.199	1.191	0.020	3.108	1.980	1.011
		200	0.132	1.002	0.001	3.150	1.996	1.009
		250	0.073	1.015	0.001	3.198	2.0161	1.006
	313	150	0.196	1.000	0.003	2.798	1.805	1.005
		200	0.152	0.999	0.013	2.857	1.824	1.002
		250	0.141	0.999	0.014	2.836	1.815	1.003
	323	150	0.055	0.999	0.037	2.705	1.701	1.006
		200	0.035	0.999	0.004	2.728	1.701	1.004
		250	0.042	0.999	0.006	2.719	1.711	0.999

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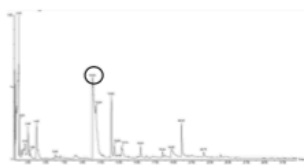
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431 **Figure 2. Analysis Result by GC-MS for the Bio-oil Produced by Cocomi Shells Pyrolysis**
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437 **Figure 3. Analysis Result by GC-MS for the Bio-oil Produced by Coffee Shells Pyrolysis**
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425 **Table 8. The Activity Coefficients were Calculated by NRTL Model**
426

Solution	Temperature	Speed	γ_a^I	γ_a^II	γ_c^I	γ_a^{II}	γ_a^{III}	γ_a^{IV}
Methanol	298	150	0.077	0.926	0.381	0.265	2.037	0.655
		200	0.064	0.482	0.337	0.001	0.039	0.065
		250	0.063	0.283	0.387	1.3×10^{-6}	0.001	7.6×10^{-7}
	313	150	4.47×10^4	0.942	0.010	8.4×10^{-7}	1.050	0.044
		200	2.71×10^6	0.986	0.056	1.5×10^{-11}	1.014	1.1×10^{-5}
		250	1.80×10^7	0.944	0.039	4.9×10^{-9}	0.880	2.4×10^{-10}
	323	150	7.92×10^4	0.980	0.010	3.6×10^{-3}	1.028	0.025
		200	3.48×10^6	0.977	0.022	4.1×10^{-14}	1.006	4.1×10^{-10}
		250	9.44×10^6	0.972	0.024	3.5×10^{-12}	0.937	1.1×10^{-9}
Acetic Acid	298	150	0.045	0.644	0.218	0.432	1.087	0.678
		200	0.039	0.331	0.210	0.015	0.361	0.552
		250	0.036	0.193	0.232	9.5×10^6	0.017	0.027
	313	150	0.000	0.734	0.037	0.036	0.995	0.551
		200	0.003	0.732	0.050	0.030	0.999	0.444
		250	0.013	0.749	0.069	0.028	1.000	0.374
	323	150	0.000	0.734	0.037	0.035	1.139	0.011
		200	0.003	0.732	0.050	0.001	1.010	0.039
		250	0.013	0.749	0.069	8.9×10^6	0.694	9.6×10^{-7}

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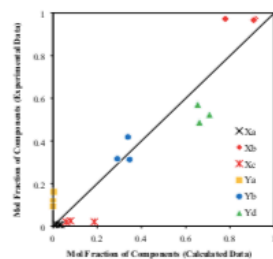
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Table 8. Sum of the Square of the Error (SSE)

Model	Temperature	SSE	
		Methanol	Acetic Acid
UNIFAC	298	0.087	0.132
	313	0.065	0.083
	323	0.053	0.084
NRTL	298	0.024	0.065
	313	0.026	0.544
	323	0.015	0.077

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440 **Figure 4. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using**
441 **Aqueous Methanol by UNIFAC Model at 298 K.**
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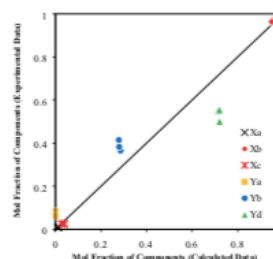
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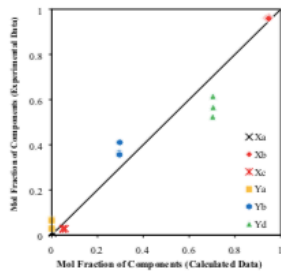
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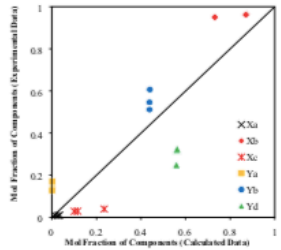
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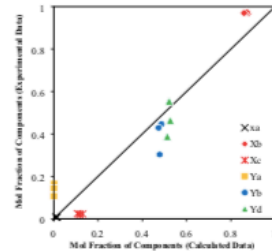
449 **Figure 5. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using**
450 **Aqueous Methanol by UNIFAC Model at 313 K.**
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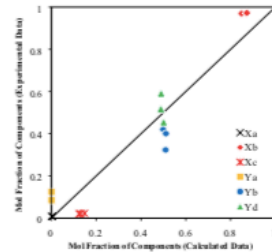
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457 **Figure 6.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
458 Aqueous Methanol by UNIFAC Model at 323 K.
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464 **Figure 7.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
465 Aqueous Acetic Acid by UNIFAC Model at 298 K.
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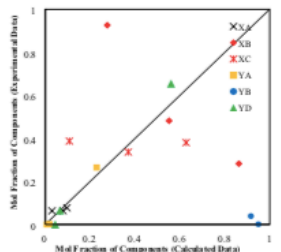
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471 **Figure 8.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
472 Aqueous Acetic Acid by UNIFAC Model at 313 K.
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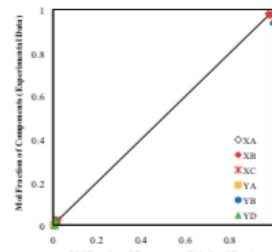
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479 **Figure 9.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
480 Aqueous Acetic Acid by UNIFAC Model at 323 K.
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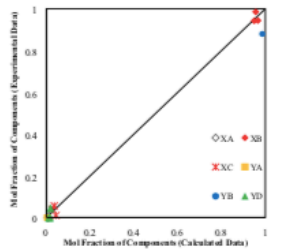
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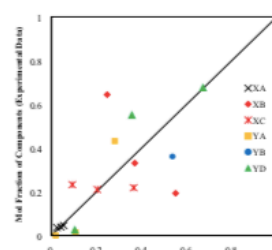
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484 **Figure 10.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
485 Aqueous Methanol by NRTL Model at 298 K.
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499 **Figure 12.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
500 Aqueous Methanol by NRTL Model at 323 K.
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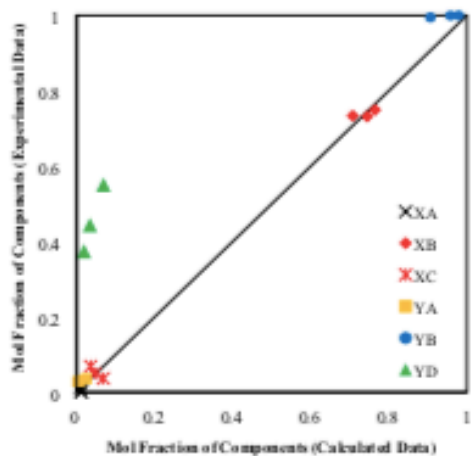
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493 **Figure 11.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
494 Aqueous Methanol by NRTL Model at 313 K.
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508 **Figure 13.** Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using
509 Aqueous Acetic Acid by NRTL Model at 298 K.
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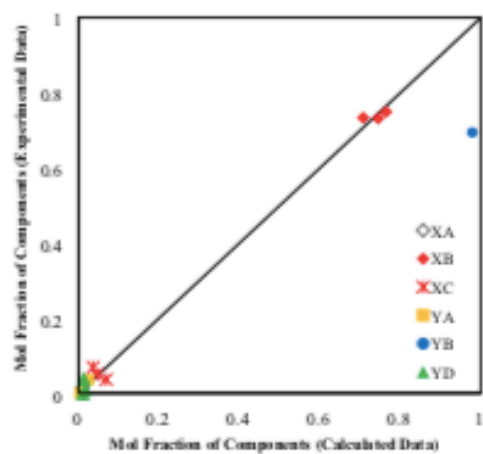
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Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 313 K



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Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Acetic Acid by NRTL Model at 323 K

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1. In general, the speed of stirring has no effect on liquid-liquid equilibrium. So the speed of stirring should not appear in Table 5 and Table 6.
2. The experimental data were still not enough to obtain accurate and credible activity coefficient.
3. In the conclusion, the author pointed out that the UNIFAC modeling provides a good correlation to liquid-liquid equilibrium data under conditions of temperature of 323 K. However, the UNIFAC model or NRTL model should be used to describe the liquid-liquid equilibrium for different temperature rather than a specific temperature.

New problems in the revised manuscript:

4. The decimal point was wrongly written as a comma in Table 6 and Table 8.
5. Two tables were labeled as Table 8 in the revised manuscript.

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Liquid phase equilibrium of phenol extraction from bio-oil produced by biomass pyrolysis using thermodynamic models

Dewi Selvia Fardhyanti, Bayu Triwibowo, Heri Istanto, Muhammad Khushi Anajib, Amalia Larasati, Windy Oktaviani



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Liquid Phase Equilibrium of Phenol Extraction from Bio-oil Produced by Biomass Pyrolysis using Thermodynamic Models

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ABSTRACT

Utilization of biomass as a new and renewable energy source is being actively conducted by various parties. One of the technology for utilizing or converting biomass as an energy source is pyrolysis, to convert biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from vapors phase of biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. The total phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). The content of phenol compounds in corrosive bio-oils still quite high, the use of this bio-oil directly will cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive, and unstable. Phenol compounds have some benefits as one of the compound for floor cleaner and disinfectant which contained in bio-oil.

The correlation between experimental data and calculations shows that the UNIQUAC Functional-group Activity Coefficients (UNIFAC) equilibrium model can be used to predict the liquid-liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the Non Random Two Liquid (NRTL) equilibrium model can be used to predict liquid-liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells.

Keywords: biomass, pyrolysis, bio-oil, UNIFAC, NRTL.

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1. INTRODUCTION

The role of fossil fuels still be crucial, but their effects will gradually be over taken by new and renewable energy sources. Along with the depletion of world oil reserves and environmental issues, various researches on renewable energy are being actively conducted by various parties. Therefore, technology needs be able to overcome the problem. One technology that can be used is pyrolysis technology where the material used comes from organic waste (biomass). Pyrolysis technology can overcome organic waste problems and make the environment healthy also convert it into value-added end product.

Biomass is an organic material derived from plants, algae, animal waste, industrial waste, forestry, and agricultural waste [1]. Biomass can be used as pyrolysis raw material to produce liquid (bio-oil), gas and solid products. Various kinds of biomass waste can be used as raw materials for the manufacture of bio-oils, such as corncobs [2], sawdust and wheat bran [3], castor [4], oak [5], durian shell [6] and coconut shell [7].

Indonesia is the third largest coffee producer in the world after Brazil and Vietnam by contributing about 6% of the world's total coffee production, also world's fourth largest coffee exporter with a market share of around 11% across the world. Coffee is one of the plantations with high economic value besides chocolate and tea. Based on data from Indonesian Ministry of Forestry and Plantation, Indonesia's coffee production in 2013 reached 691,160 tons with a plantation area of 47,800 Ha [8]. Waste from coffee processing that abundant and not utilized optimally is the coffee shell. Large waste of coffee shells derived from plantations was approximately 11.08% which potentially cause environment hazard. Coffee shells are a waste that has high lignocellulose content which have potential to be used as bio-oil. Bio-oil is a blackish liquid fuel derived from biomass such as wood, coffee shells and other biomass. Bio-oil contains lignin components (23.4%), cellulose (34.2%) and hemicellulose (24.5%) [9].

In Indonesia, in addition to the coffee shell, one of the available biomass resources is coconut shell. Availability of coconut shell can be seen from the production of coconut fruit in Indonesia. Coconut is one of Indonesia's main commodities with an average production of 3,000,000 tons / year [10]. The utilization of coconut is very diverse, but for the utilization and way of handling waste is still limited. Coconut fruit can be used as food in daily life for the purpose of consumed directly. Coconut shells nowadays, lots of it become garbage that can not be utilized. One of the waste handling is to utilize coconut shell waste as a base material in the manufacture of bio-oil. Characterization studies on coconut shells have been done through proximate, ultimate, and Thermo Gravimetric (TG) analyzes.

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The results of this analysis indicate that coconut shell has a high potential to produce fuel fluid by pyrolysis conversion process [11]. Chemical content of coconut shell is cellulose (34%), hemicellulose (21%), lignin (27%) and ash (18%) [12].

Before converted into bio-oil, coconut shell waste and coffee shell need to be treated first. It takes a process before converting the waste of coffee shell into bio-oil which is pyrolysis. Pyrolysis is the process of decomposition of a substance or material that is done at a relatively high temperature. Pyrolysis results in the form of gas, liquid and solid. The solid result of this process is char (char). Gas and liquid products can be used as a source of bio-oil. Bio-oil can be used as an alternative fuel that is easier to store and distribute than biomass in solid form. Bio-oil made from coconut shell material using slow pyrolysis with pyrolysis temperature 250-300 °C produces liquid which very acidic and corrosive. The largest organic component in bio-oil is the lignin derivative of phenol, alcohol, organic acids and carbonyl compounds such as ketones, aldehydes and esters [13]. The nature of the acid is caused by the content of the dominant phenol compounds.

Phenol compounds in addition to causing the smell of smoke in bio-oil, also showed antioxidant activity and corrosive [14]. Therefore, the use of this bio-oil will directly cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive and unstable [6].

Phenol needs to be extracted from bio-oil to reduce its corrosive properties. In addition, phenol is a compound that has a high economic value because of its expensive and widely used. Phenols have many important uses such as disinfectants, resins, pesticides, explosives, and drugs. The method that can be used to extract phenol compounds from bio-oil is by liquid-liquid extraction method using methanol solvent. This has been done by Jabrinsk [15] on the isolation of phenol compounds from bio-oils from pyrolysis of forest waste and Mantilla [16] on the extraction of phenol compounds from bio-oils from pyrolysis of agricultural waste. Dahlia [17] conducted research on liquid-liquid extraction of phenol compound from bio-oil of pyrolysis resulted of palm oil empty bunch using methanol solvent. The yield of phenol resulting from this extraction is 40%. In this study, it is expected that the liquid-liquid extraction method can be used to separate the phenol compounds from coconut shell pyrolysis bio-oils. The purpose of phenol extraction is to improve the quality of bio-oil and reduce the corrosivity, so when used as fuel, it will cause no damage to the engine.

Biomass pyrolysis converted to bio-oil studies have been conducted. Bio-oil produced from the pyrolysis process can be used as fuel for engines, turbines and boilers. The bio-oil which processed further by liquid-liquid extraction process will reduce the corrosive nature of bio-oil and obtained chemicals which one of them is phenol. Research on extraction of phenol compounds from bio-oil is still very limited. In addition, thermodynamic modelling study on liquid-liquid equilibrium of phenol

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extraction process from bio-oil still needs to be investigated further.

To design extraction columns required various information such as liquid-liquid equilibrium data (Liquid-liquid Equilibrium). Thermodynamic models such as Equation of State or Activity Coefficient are used to correlate the experimental data to obtain optimal interaction parameters. Optimal parameters can be used to predict liquid-liquid equilibrium data to produce an equilibrium graph that is used to design extraction columns [18]. The study on liquid-liquid equilibrium include NRTL, e-NRTL, UNIQUAC, and UNIFAC.

Given the importance of thermodynamic modeling in this liquid-liquid extraction process and the price and benefits of phenol in the industrial world, this research becomes an important thing to do. From the equilibrium data, the optimum condition will be obtained where the highest yield of extract obtained in the extraction process of phenol compound from bio-oil from coconut pyrolysis. Liquid-liquid equilibrium data of the components of the system can be used in the design of the equipment in separation process.

2. EXPERIMENTAL SECTION

2.1 Chemicals

All reagents used here are A.R. grade and used as received without further purification. Methanol, acetic acid, and chloroform were purchased from e-Merck (Germany). The distilled water is available at the laboratory. Coffee shell and coconut shell were obtained from public market in Semarang.

2.2 Equilibrium Procedure

Instruments used in this study:

- Shaker bath, product by Memmert WB14, SV1422, Schwabach, Germany.
- Gas Chromatography (GC), product by GC 6820 Agilent Technologies, column: Rastek RXI-5MS002E.
- Gas Chromatography-Mass Spectroscopy (GC-MS), product by GCMS-QP2010S Shimadzu, column: Rastek RNg-5MS.

The material systems and experimental conditions for the equilibrium extraction are summarized in Table 1.

Table 1. Material Systems and Conditions for Equilibrium Extraction

The bio-oil feed solution that containing phenol was extracted using two-stage extraction. The first stage uses distilled water as a polar solvent and chloroform as a non-polar solvent. Extraction was conducted with ice-bath method at 4-5°C, 200 rpm and stirring time for 2 hours. The extracts obtained

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at this first stage are used as feeds on the second stage extraction. At the second stage extraction, each extract was extracted using a methanol solvent under operating conditions shown in Table 1.

2.3 Analysis and Characterization

After equilibrium attained (60 minutes), the mixtures were poured into a separating funnel, settled for an hour and separated into two phases. Then the two phases were weighed. The extract phase and the raffinate phase of each extraction processes are analyzed by GC to obtain the concentration of each component. The principal conditions of this analysis are shown in Table 2.

Table 2. Conditions of Analysis Using GC

3. RESULTS AND DISCUSSION

3.1 Bio-Oil Content

The chromatograms of the GC-MS analysis are shown in Fig. 1 and 2. The results of the GC-MS analysis of the bio-oil showed that they contained more than six chemical compounds such as ethyl ester, phenol, furfural, acetic acid, and others.

Figure 1. The Chromatogram of Bio-Oil Produced from Coconut Shell Pyrolysis

Table 3. The Components of Bio-oil Produced from Coconut Shell Pyrolysis

Figure 2. The Chromatogram of Bio-Oil Produced from Coffee Shell Pyrolysis

Table 4. Component of Bio-oil Produced from Coffee Shell Pyrolysis

Table 3 and 4 represent that bio-oil contains more than 6 main components. The percentage (%) of the compositions of the bio-oil analysis by GC-MS was obtained from the percentage area of the peak or the height of the peak in the chromatograms. Using a method of GC-MS, this work presents a composition of the bio-oil generated in the experiment. The total phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). It was caused by the pyrolysis processing at 300-350°C (low temperature carbonization) that produce more substances contain ester and phenolic compounds. The composition of biomass such as hemicellulose, cellulose and lignin will be oxidized to phenol as the main component of the bio-oil.

3.2 Equilibrium data on phenol extraction of bio-oil from coconut pyrolysis results

In this research the variables studied were temperature, speed of stirring and type of solvents (aqueous methanol and aqueous acetic acid). Generally, all extracts are rich of phenol but the mole fraction significantly depends on extraction conditions: temperature, speed of stirring and type of

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solvents. Methanol and acetic acid need to be mixed with water because their polarities are not strong enough to form immiscible mixture with bio-oil [19].

The mole fractions of any components in the raffinate phase and the extract phase are constant after 60 minutes, so the system is assumed to be in equilibrium. The amount of water transferred from the solvent into the raffinate phase was negligible [19].

In this work, we are reported experimental data for the phenol extraction from bio-oil produced from coconut shell and the phenol extraction from bio-oil produced from coffee shell.

Table 5 and 6 show the equilibrium of multi-components extraction data for aqueous acetone and aqueous methanol solvent in the extract phase and raffinate phase.

Table 5. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Methanol from Bio-oil Produced from Coconut Shell.

Table 6. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Acetic Acid from Bio-oil Produced from Coffee Shell.

3.3 Liquid-liquid Equilibrium for The Phenol Extraction Process using UNIFAC dan NRTL Models.

The UNIFAC and NRTL models were used to correlate the experimental data obtained for the systems under study. UNIFAC equation and NRTL equation (Smith, 1996) were used to correlate experimental phase equilibrium data. The activity coefficient models applied in this study are as follows:

- UNIFAC: [20]

UNIFAC is a UNIQUAC Functional-group Activity Coefficients model based on the UNIQUAC equation by using functional groups of a molecule contained in the mixture to calculate the activity coefficients. This model was developed by Fredenslund, Jones, and Prausnitz (1975). The UNIFAC modeling is a continuation of the UNIQUAC modeling development. This model has the advantage of performing phase equilibrium estimation even in the absence of experimental data. This thermodynamic model is used to predict the estimation of non-electrolyte activity in non-ideal mixtures. The UNIFAC model calculates the activity coefficients by using functional groups present in the molecules that form the liquid mixture. If the value of the component activity coefficient has a value close to one, this indicates that the component is in the ideal state in the system [21]. The correlation in the UNIFAC model is used in order to reduce the number of binary interactions measured to predict the state of the system. The correlation for the UNIFAC model is shown as follows:

$$\ln \gamma_i = \ln \gamma_i^c + \ln \gamma_i^R \quad (1)$$

$$\ln \gamma_i^c = 1 - J_i + \ln J_i - 5q_i \left(1 - \frac{L_i}{L_j} + \ln \frac{L_i}{L_j} \right) \quad (2)$$

$$\ln \gamma_i^R = q_i \left[1 - \sum_k \left(\theta_k \frac{r_{ik}}{r_k} - e_{ik} \ln \frac{r_{ik}}{r_k} \right) \right] \quad (3)$$

$$J_i = \frac{-z_{ij}}{\sum_j r_j \sigma_j} \quad (4)$$

$$L_i = \frac{q_i z_i}{\sum_j r_j \sigma_j} \quad (5)$$

$$r_i = \sum_k v_k^{(i)} R_k \quad (6)$$

$$q_i = \sum_k v_k^{(i)} Q_k \quad (7)$$

$$e_{ik} = \frac{z_{ik}^{(i)}}{q_i} \quad (8)$$

$$\beta_{ik} = \sum_m \sigma_m \tau_{mk} \quad (9)$$

$$\theta_k = \frac{\sum_i \sigma_i \tau_{ik}}{\sum_i r_i \sigma_i} \quad (10)$$

$$\tau_{mk} = \sum_n \theta_n \tau_{mnk} \quad (11)$$

$$\tau_{mnk} = \exp \frac{-z_{mnk}}{T} \quad (12)$$

Where:

- $\ln \gamma_i^c$ = Combinatorial component
- $\ln \gamma_i^R$ = residual component
- J_i = component surface area component (i)
- L_i = component volume fraction (i)
- r_i = pure component surface area parameter (i)
- x_i = pure component mol fraction (i)
- q_i = pure component volume parameter (i)
- $v_k^{(i)}$ = number of sub-group (k) on component (i)
- R_k = parameter area for sub-group (k) ($R_k = A_{kk}/(2.5 \times 10^3)$, where A_{kk} is number of sub-group van der Waals surface area)
- Q_k = number of parameter volume subgroup (k) ($Q_k = V_{kk}/15.17$, where V_{kk} is number of Van der Waals volume subgroup)
- τ_{mk} = group of interaction parameter from group m and k [18]
- x_i = liquid mol fraction of component (i)

UNIFAC equation can be obtained to :

$$\ln \gamma_i = \ln \left[1 - J_i + \ln J_i + 5q_i \left(1 - \frac{L_i}{L_j} + \ln \frac{L_i}{L_j} \right) \right] + \ln \left\{ \left[1 - \sum_k \left(\frac{\theta_k \sigma_k \tau_{ik}}{\sum_n \theta_n \tau_{in}} - \frac{r_k^{(i)} Q_k}{q_i} \ln \frac{\theta_k \sigma_k \tau_{ik}}{\sum_n \theta_n \tau_{in}} \right) \right] \right\} \quad (13)$$

8

b) NRTL: [20]

Non Random Two Liquid (NRTL) is derived from the Scott two liquid model and it is assumed that randomness is the same as the model used on the Wilson model. The NRTL model has a non-random parameter (α_{12}) making it possible to use in a wide variety of mixtures. The NRTL model uses only binary parameters to calculate the phase equilibrium properties.

The NRTL (Non Random Two-Liquid) equation was first developed by Renon and Prausnitz (1968). The NRTL model is derived from the Scott two liquid model and it is assumed that the non-random parameter is the same as the model used in the Wilson Model. The NRTL equation is developed based on a two-fluid theory with the addition of a non-random parameter (α), which makes the equation applicable to different types of liquid-liquid mixture and equilibrium by choosing the appropriate α value. The parameter estimation for this equation is done by setting the α value at a certain value and subsequently optimizing the objective function to obtain two optimal parameter interactions. The recommended α value is 0.2 - 0.47. The value of α is often adjusted to certain conditions, but usually the value of $\alpha = 0.3$. Unlike the Wilson equation which contains two adjustable parameters, the NRTL equation contains three parameters, so the NRTL equation can be an alternative model if other models are inadequate. This equation can also estimate the equilibrium of multicomponent systems without additional parameters.

The NRTL equation can represent liquid-liquid equilibrium for both binary and multicomponent systems only with binary parameters. The equation is simpler than the UNIQUAC equation, but it has disadvantage because it contains three parameters for each pair of components. The third parameter (α) is estimated based on the state of the component and sometimes for certain substances the extreme number is obtained. The NRTL equation is expressed:

$$\frac{G^E}{RT} = \sum_{i=1}^n x_i \sum_{j=1}^n \tau_{ij} \frac{x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} \quad (14)$$

$$\tau_{ij} = \frac{B_{ij} - B_{ji}}{RT} \quad (15)$$

$$B_{ij} = B_{ji} = 0 \quad (16)$$

$$G_{ij} = \exp(-\alpha_{ij} \tau_{ij}) \quad (17)$$

$$\alpha_{ij} = \alpha_j \quad (18)$$

$$\ln \gamma_i = \frac{\sum_{j=1}^n x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} + \sum_{j=1}^n x_j \frac{x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} \tau_{ij} - \sum_{k=1}^n \frac{x_k G_{ik}}{\sum_{l=1}^n x_l G_{il}} \quad (19)$$

$$x_i = \exp \left(\frac{\sum_{j=1}^n x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} + \sum_{j=1}^n x_j \frac{x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} \tau_{ij} - \sum_{k=1}^n \frac{x_k G_{ik}}{\sum_{l=1}^n x_l G_{il}} \right) \quad (20)$$

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$$V_i = \exp \left(\frac{\sum_{j=1}^n x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} + \sum_{j=1}^n x_j \frac{x_j G_{ij}}{\sum_{k=1}^n x_k G_{ik}} \tau_{ij} - \sum_{k=1}^n \frac{x_k G_{ik}}{\sum_{l=1}^n x_l G_{il}} \right) \quad (21)$$

Where:

 G^E = molar Gibbs energy R = gas constant T = temperature x = liquid phase mol fraction i, j, k = component of 1, 2, 3, ..., n α = non-random parameter for NRTL g, G = component parameter γ = activity coefficient τ = empirical constant for NRTL**Table 7. The Activity Coefficients were Calculated by UNIFAC Model.****Table 8. The Activity Coefficients were Calculated by NRTL Model.**

Table 7 and 8 shows the value of activity coefficient on phenol extraction using methanol solvent obtained from calculation with UNIFAC and NRTL modeling. From coefficient value of activity on each component shows that methanol and acetic acid component in the extract phase is in ideal condition because the value of activity coefficient obtained close to one. The activity coefficient value of phenol and water in the extract phase is not close to one, it means that phenol and water components are far from ideal conditions for the system. The ideal conditions of the components in the system show that the component is dominant, so the properties of the mixture are closer to the dominant component properties either in the extract or raffinate phase. In the raffinate phase, it is known that the bio-oil has a coefficient of activity close to the one, indicating that the bio-oil is in ideal conditions for the system, while the phenol and solvent (acetic acid or methanol) are far from ideal conditions. So non-ideality studies are very important for those systems [21].

Figure 3. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 298 K.

Where:

 x_e : mole fraction of component in extract phase x_s : mole fraction of solvent in extract phase x_w : mole fraction of water in extract phase y_r : mole fraction of component in raffinate phase

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 y_i : mole fraction of solvent in raffinate phase y_o : mole fraction of bio-oil in raffinate phase

Figure 4. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 313 K.

Figure 5. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 323 K.

Figure 6. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 298 K.

Figure 7. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 313 K.

Figure 8. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 323 K.

Figure 9. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 298 K.

Figure 10. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 313 K.

Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 323 K.

Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 298 K.

Figure 13. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 313 K.

Figure 14. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Acetic Acid by NRTL Model at 323 K.

Where:

 y_e^i : mole fraction of component in extract phase y_e^s : mole fraction of solvent in extract phase y_e^w : mole fraction of water in extract phase y_r^i : mole fraction of component in raffinate phase y_r^s : mole fraction of solvent in raffinate phase y_r^o : mole fraction of bio-oil in raffinate phase

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The result of correlation of liquid-liquid equilibrium data by using UNIFAC and NRTL modeling gives good agreement between experimental data and calculation. This is shown by the value of SSE (Sum of the Square of the Error) is very small. The SSE values in UNIFAC and NRTL modeling can be seen in Table 9. The value of SSE is 0,015 - 0,554.

Table 9. Sum of the Square of the Error (SSE)

Based on Table 9, it can be seen that extraction with methanol solvent for NRTL model has the smallest SSE value compared to the extraction with acetic acid solvent for UNIFAC model. The smaller the SSE values obtained show that the correlation between the experimental data and the calculation data indicates a better correlation. Good correlation results show that equilibrium modeling of UNIFAC and NRTL can be used to predict liquid-liquid equilibrium in phenol extraction process from cocunut shell pyrolysis bio-oil and coffee shell using methanol solvent and acetic acid.

Figure 3-14 shows that the visual comparison of calculated and experimental mole fractions in the extract phase and the raffinate phase for the methanol solvent system by NRTL model are better than the other conditions (by UNIFAC model and acetic acid solvent).

4. CONCLUSION

NRTL modeling gives a good correlation to liquid-liquid equilibrium data for ternary system on phenol extraction of bio-oil using methanol solvent compared to the UNIFAC modeling using acetic acid solvent.

The activity coefficients of water and phenol in their phase are far from unity. It means that they are far from ideal condition. The activity coefficients of methanol, acetic acid, and bio-oil in their phases are close to one. It means that they are nearly ideal in their phases.

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Table 1. Material Systems and Conditions for Equilibrium Extraction.

	Quantity
feed	bio-oil
feed volume	2.5 mL
solvent	aqueous methanol
	aqueous acetic acid
mole fraction of water in solvent	0.2
solvent-feed mass ratio	1
shaking time	60 minutes
amplitude of shaking	150, 200, 250 rpm
temperature	298, 313, 323 K.

Table 2. Conditions of Analysis Using GC

Column: Rastek RXI-5MS		
Column:		
Inner diameter	[m]	3.2×10^{-4}
Length	[m]	30
Carrier gas		He
Split ratio	[-]	153
Flow rate	[cm ³ /sec]	26.6
Sample volume	m ³	1.10^6
Injection temperature	[K]	553
Column temperature	[K]	313 - 573
Pressure column	kPa	10.0
Column flow	ml./min	0.54
Detector (FID) temperature	[K]	573

Table 3. The Main Components of Bio-oil Produced by Pyrolysis of Coconut Shells

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Peak	Component	Percentage
1	Ethyl ester	37.60
2	Formic acid	4.02
3	1-Hydroxy-2-butanone	3.89
4	Furfural	5.45
5	Phenol	40.01
7	2-Methoxy-phenol	7.02

Table 4. The Main Components of Bio-oil Produced by Pyrolysis of Coffee Shells

Peak	Component	Percentage
1	Urea	5%
2	Acetic acid	26%
5	Furfural	6%
6	Phenol	34%
7	Phenol, 2-methoxy-	7%
8	Phenol, 2,6-dimethoxy	4%

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Table 5. Equilibrium of Components Extraction Data for Phenol Extraction using Aqueous Methanol from Bio-oil Produced from Coconut Shell and Coffee Shell.

Temperature	Biomass	X_1	X_2	X_3	Y_1	Y_2	Y_3
298 K	Coconut Shell	0.096	0.876	0.363	0.229	0.921	0.560
		0.078	0.852	0.370	0.219	0.916	0.653
		0.030	0.863	0.311	0.165	0.950	0.544
	Coffee Shell	0.076	0.926	0.333	0.259	0.891	0.540
		0.058	0.902	0.340	0.249	0.886	0.633
		0.016	0.913	0.281	0.195	0.920	0.524
313 K	Coconut Shell	0.077	0.949	0.043	0.085	0.549	0.365
		0.083	0.956	0.035	0.086	0.414	0.501
		0.071	0.967	0.026	0.062	0.555	0.383
	Coffee Shell	0.087	0.956	0.023	0.365	0.975	0.018
		0.093	0.963	0.025	0.343	0.982	0.014
		0.081	0.974	0.016	0.292	0.986	0.012
323 K	Coconut Shell	0.054	0.972	0.023	0.395	0.945	0.025
		0.051	0.978	0.017	0.373	0.952	0.021
		0.049	0.980	0.015	0.322	0.956	0.019
	Coffee Shell	0.044	0.922	0.033	0.235	0.988	0.010
		0.041	0.928	0.030	0.287	0.991	0.018
		0.039	0.930	0.026	0.343	0.992	0.017

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Table 7. The Activity Coefficients were Calculated by UNIFAC Model.

Solution	Temperature	Biomass	γ_1^L	γ_2^L	γ_3^L	γ_1^H	γ_2^H	γ_3^H
Methanol	298 K	Coconut Shell	0.176	1.989	0.037	0.005	0.063	2.360
		Coffee Shell	0.032	1.994	0.004	0.008	0.098	2.398
		Coconut Shell	0.019	1.999	0.003	0.012	0.141	2.435
		Coffee Shell	0.044	1.883	0.067	0.014	0.047	3.112
		Coconut Shell	0.027	1.976	0.019	0.022	0.981	3.309
		Coffee Shell	0.149	1.789	0.006	0.028	0.035	2.143
Methanol	313 K	Coconut Shell	0.119	1.791	0.005	0.027	0.070	2.215
		Coffee Shell	0.112	1.792	0.003	0.034	0.085	2.191
		Coconut Shell	0.157	1.965	0.042	0.023	0.032	2.143
		Coffee Shell	0.161	1.983	0.029	0.025	0.036	2.215
		Coconut Shell	0.178	1.992	0.081	0.025	0.062	2.191
		Coffee Shell	0.049	1.788	0.007	0.017	0.054	2.181
Methanol	323 K	Coconut Shell	0.028	1.831	0.001	0.017	0.052	2.222
		Coffee Shell	0.034	1.830	0.001	0.017	0.052	2.191
		Coconut Shell	0.058	1.839	0.012	0.021	0.041	2.181
		Coffee Shell	0.053	1.780	0.021	0.036	0.064	2.222
		Coconut Shell	0.088	1.832	0.028	0.032	0.068	2.191
		Coffee Shell	0.199	1.191	0.020	0.023	0.078	3.108
Acetic Acid	298 K	Coconut Shell	0.132	1.002	0.018	0.035	0.081	3.150
		Coffee Shell	0.075	1.015	0.015	0.044	0.086	3.198
		Coconut Shell	0.206	1.222	0.033	0.022	0.083	3.108
		Coffee Shell	0.325	1.123	0.042	0.043	0.087	3.150
		Coconut Shell	0.368	1.015	0.047	0.078	0.091	3.198
		Coffee Shell	0.396	1.023	0.003	0.038	0.081	2.798
Acetic Acid	313 K	Coconut Shell	0.152	1.319	0.013	0.040	0.087	2.857
		Coffee Shell	0.141	1.439	0.014	0.043	0.091	2.836
		Coconut Shell	0.211	1.032	0.033	0.056	0.108	2.798
		Coffee Shell	0.278	1.804	0.037	0.069	0.115	2.857
		Coconut Shell	0.305	1.829	0.044	0.053	0.130	2.836
		Coffee Shell	0.055	1.925	0.037	0.044	0.071	2.705
Acetic Acid	323 K	Coconut Shell	0.082	2.009	0.006	0.044	0.083	2.719
		Coffee Shell	0.087	1.909	0.006	0.063	0.111	2.705
		Coconut Shell	0.087	1.921	0.067	0.067	0.135	2.728
		Coffee Shell	0.103	2.001	0.070	0.051	0.135	2.719

Table 6. Equilibrium of Components Extraction Data for Phenol Extraction using Acetic Acid from Bio-oil Produced from Coconut Shell and Coffee Shell.

Temperature	Biomass	X_1	X_2	X_3	Y_1	Y_2	Y_3
298 K	Coconut Shell	0.024	0.247	0.365	0.028	0.470	0.672
		0.024	0.370	0.204	0.011	0.537	0.457
		0.024	0.551	0.093	0.019	0.878	0.104
	Coffee Shell	0.033	0.707	0.375	0.033	0.431	0.687
		0.031	0.744	0.214	0.046	0.587	0.472
		0.030	0.764	0.103	0.022	0.832	0.119
313 K	Coconut Shell	0.012	0.707	0.068	0.025	0.907	0.687
		0.011	0.744	0.047	0.008	0.958	0.339
		0.010	0.764	0.036	0.003	0.979	0.019
	Coffee Shell	0.021	0.217	0.078	0.035	0.912	0.702
		0.021	0.340	0.057	0.003	0.963	0.354
		0.020	0.521	0.046	0.002	0.955	0.034
323 K	Coconut Shell	0.042	0.677	0.068	0.025	0.907	0.016
		0.011	0.714	0.047	0.008	0.958	0.012
		0.010	0.734	0.036	0.003	0.979	0.011
	Coffee Shell	0.022	0.677	0.078	0.030	0.896	0.031
		0.021	0.714	0.057	0.005	0.931	0.027
		0.023	0.734	0.046	0.008	0.944	0.026

Table 8. The Activity Coefficients were Calculated by NRTL Model.

Solution	Temperature	Biomass	γ_1^L	γ_2^L	γ_3^L	γ_1^H	γ_2^H	γ_3^H
Methanol	298 K	Coconut Shell	0.077	0.996	0.381	0.005	0.218	2.037
		Coffee Shell	0.064	0.982	0.337	0.001	0.222	2.037
		Coconut Shell	0.063	0.988	0.387	1.3e10 ⁻⁶	0.245	2.045
		Coffee Shell	0.132	1.003	0.289	0.001	0.468	1.803
		Coconut Shell	0.157	1.045	0.376	0.001	0.458	1.823
		Coffee Shell	0.221	1.067	0.401	0.001	0.312	2.007
Methanol	313 K	Coconut Shell	4.47e10 ⁻⁶	1.003	0.010	8.4e10 ⁻⁶	0.278	1.050
		Coffee Shell	2.71e10 ⁻⁶	1.009	0.056	1.5e10 ⁻⁶	0.308	1.014
		Coconut Shell	1.80e10 ⁻⁶	1.023	0.039	4.9e10 ⁻⁶	0.347	0.880
		Coffee Shell	0.004	1.000	0.067	0.007	0.280	0.999
		Coconut Shell	0.005	0.997	0.069	0.003	0.480	1.012
		Coffee Shell	0.007	1.015	0.306	0.020	0.308	1.003
Methanol	323 K	Coconut Shell	7.92e10 ⁻⁶	1.110	0.010	3.0e10 ⁻⁶	0.113	1.028
		Coffee Shell	3.48e10 ⁻⁶	1.088	0.022	4.1e10 ⁻⁶	0.132	1.006
		Coconut Shell	9.44e10 ⁻⁶	1.122	0.024	3.5e10 ⁻⁶	0.133	0.937
		Coffee Shell	0.001	1.313	0.054	0.004	0.167	1.012
		Coconut Shell	0.008	1.402	0.057	0.007	0.175	1.097
		Coffee Shell	0.008	1.399	0.055	0.003	0.175	1.032
Acetic Acid	298 K	Coconut Shell	0.045	1.209	0.218	0.002	0.210	1.087
		Coffee Shell	0.039	1.211	0.110	0.015	0.200	1.265
		Coconut Shell	0.036	1.309	0.232	9.5e10 ⁻⁶	0.200	1.478
		Coffee Shell	0.146	1.448	0.087	0.057	0.214	2.014
		Coconut Shell	0.135	1.443	0.075	0.039	0.247	2.004
		Coffee Shell	0.128	1.761	0.102	0.078	0.211	2.065
Acetic Acid	313 K	Coconut Shell	0.002	2.003	0.037	0.036	0.126	0.995
		Coffee Shell	0.003	2.032	0.050	0.030	0.119	0.999
		Coconut Shell	0.013	1.956	0.069	0.028	0.117	1.000
		Coffee Shell	0.019	1.782	0.023	0.022	0.209	1.119
		Coconut Shell	0.018	1.834	0.026	0.025	0.211	1.118
		Coffee Shell	0.021	2.000	0.080	0.025	0.211	1.097
Acetic Acid	323 K	Coconut Shell	0.115	1.546	0.037	0.035	0.308	1.139
		Coffee Shell	0.138	1.690	0.050	0.061	0.311	1.010
		Coconut Shell	0.200	1.800	0.069	8.9e10 ⁻⁶	0.289	1.021
		Coffee Shell	0.054	1.906	0.046	0.013	0.267	1.290
		Coconut Shell	0.090	2.035	0.067	0.016	0.214	1.317
		Coffee Shell	0.093	2.088	0.099	0.021	0.222	1.320

Table 9. Sum of the Square of the Error (SSE)

Model	Temperature	SSE	
		Methanol	Acetic Acid
UNIFAC	298	0.087	0.132
	313	0.065	0.083
	323	0.053	0.084
NRTL	298	0.024	0.065
	313	0.026	0.054
	323	0.015	0.077

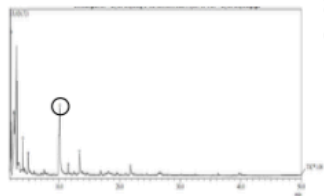


Figure 1. Analysis Result by GC-MS for the Bio-oil Produced by Coconut Shells Pyrolysis



Figure 2. Analysis Result by GC-MS for the Bio-oil Produced by Coffee Shells Pyrolysis

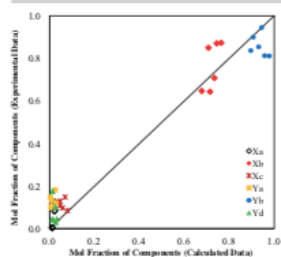


Figure 5. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 323 K.

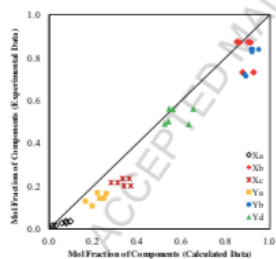


Figure 6. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 298 K.

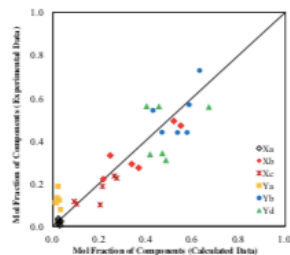


Figure 3. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 298 K.

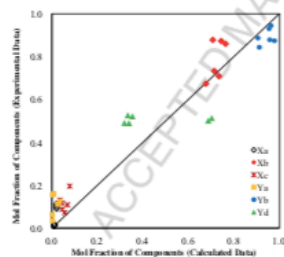


Figure 4. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by UNIFAC Model at 313 K.

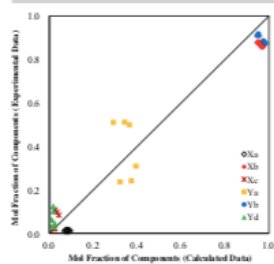


Figure 7. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 313 K.

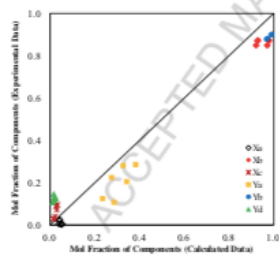


Figure 8. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by UNIFAC Model at 323 K.

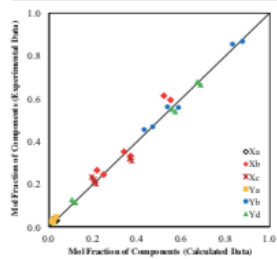


Figure 9. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 298 K

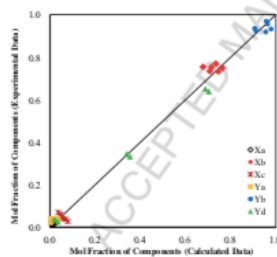


Figure 10. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 313 K

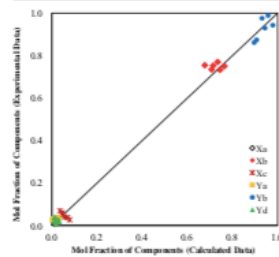


Figure 11. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Methanol by NRTL Model at 323 K

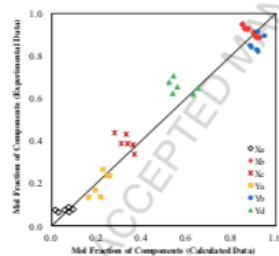


Figure 12. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 298 K

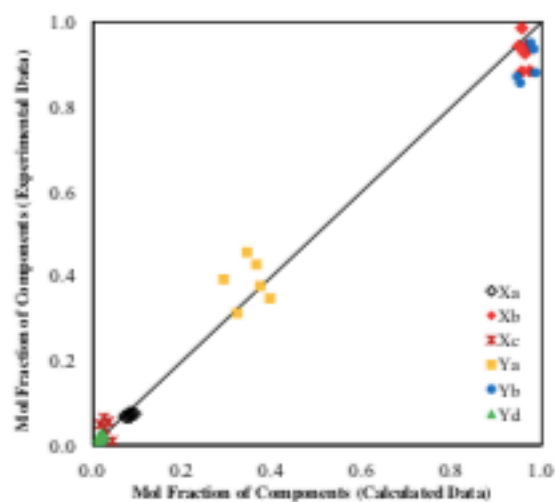


Figure 13. Comparison of Calculated and Experimental Mole Fractions of the Phenol Extraction using Aqueous Acetic Acid by NRTL Model at 313 K

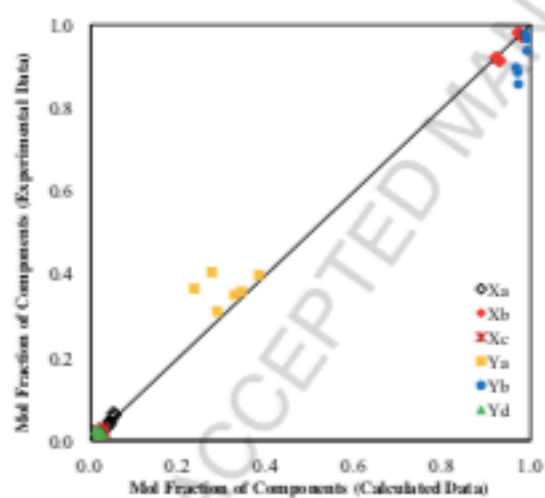



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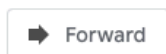
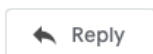
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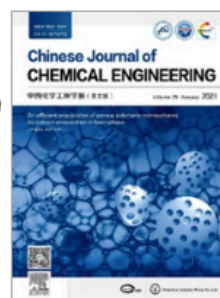


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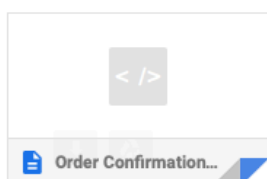
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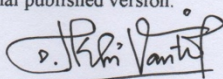
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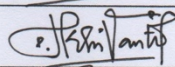
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Abstract

Utilization of biomass as a new and renewable energy source is being actively conducted by various parties. One of the technology for utilizing or converting biomass as an energy source is pyrolysis, to convert biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from vapors phase of biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. The total phenolic compounds contained in bio-oil is 47.03% (coconut shell) and 45% (coffee shell). The content of phenol compounds in corrosive bio-oils still quite high, the use of this bio-oil directly will cause various difficulties in the combustion system due to high viscosity, low calorific value, corrosive, and unstable. Phenol compounds have some benefits as one of the compound for floor cleaner and disinfectant which contained in bio-oil.

The correlation between experimental data and calculations shows that the UNIQUAC Functional-group Activity Coefficients (UNIFAC) equilibrium model can be used to predict the liquid–liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the Non Random Two Liquid (NRTL) equilibrium model can be used to predict liquid–liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells.

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Liquid phase equilibrium of phenol extraction from bio-oil produced by biomass pyrolysis using thermodynamic models

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ABSTRACT

Utilization of biomass as a new and renewable energy source is being actively conducted by various parties. One of the technologies for utilizing or converting biomass as an energy source is pyrolysis, to convert biomass into a more valuable product which is bio-oil. Bio-oil is a condensed liquid from the vapor phase of biomass pyrolysis such as coconut shells and coffee shells. Biomass composition consisting of hemicellulose, cellulose, and lignin will oxidize to phenol which is the main content in bio-oil. The total phenolic compound contained in bio-oil are 47.02% (coconut shell) and 43% (coffee shell). The content of phenol compounds in coconut bio-oils still quite high, the use of the bio-oil directly will cause various difficulties in the combustion system due to high viscosity, low calorific value, moisture, and instability. Phenol compounds have some benefits as one of the compounds for cleaners and disinfectants which are contained in bio-oil.

The correlation between experimental data and calculation shows that the UNIFAC Functional-group Activity Coefficients (UNIFAC) equilibrium model can be used to predict the liquid–liquid equilibrium in the phenol extraction process of the coconut shell pyrolysis bio-oil. While the Non-Random Two-Liquid (NRTL) equilibrium model can be used to predict liquid–liquid equilibrium in the extraction process of phenol from bio-oil pyrolysis of coffee shells.

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1. Introduction

The use of fossil fuel is still crucial, but their reserves will gradually be over taken by new and renewable energy sources. Along with the depletion of world oil reserves and environmental issues, various researches on renewable energy are being actively conducted by various parties. Therefore, technology needs to be able to overcome the problem. One technology that can be used is pyrolysis technology where the material used comes from organic waste (biomass). Pyrolysis technology can overcome organic waste problems and make the environment healthy by converting waste into value-added end product.

Biomass is an organic material derived from plants, algae, animal waste, industrial waste, forestry, and agricultural waste [1]. Biomass can be used as pyrolysis raw material to produce liquid (bio-oil), gas and solid products. Various kinds of biomass waste can be used as raw materials for the manufacture of bio-oils, such as coconuts [2], sawdust and wheat bran [3], castor [4], oak [5], durian shell [6] and coconut shell [7].

Indonesia is the third largest coffee producer in the world after Brazil and Vietnam by contributing about 6% of the world's total coffee

production, also world's fourth largest coffee exporter with a market share of around 11% across the world. Coffee is one of the plantations with high economic value besides chocolate and tea. Based on data from the Indonesian Ministry of Forestry and Plantation, Indonesia's coffee production in 2013 reached 691,160 tons with a plantation area of 47800 Ha [8]. Waste from coffee processing that is abundant and not utilized optimally is the coffee shell. A large percentage of waste from coffee shells derived from plantations amounts to approximately 11.08% which potentially causes environment hazard. Coffee shells are a waste that has high lignocellulose content which has a potential to be used as bio-oil. Bio-oil is a blackish liquid fuel derived from biomass such as wood, coffee shells and other biomass. Bio-oil contains lignin components (21.4%), cellulose (34.2%) and hemicellulose (24.5%) [8].

In Indonesia, in addition to the coffee shell, one of the available biomass resources is coconut shell. Availability of coconut shell can be seen from the production of coconut fruit in Indonesia. Coconut is one of Indonesia's main commodities with an average production of 300000 tons per year [9]. The utilization of coconut is very diverse, but the utilization and way of handling waste are still limited. Coconut fruit can be used as food in daily life for the purpose of direct consumption. Nowadays, lots of coconut shells become garbage that cannot be utilized. One of the waste handling is to utilize coconut shell waste as a base material in the manufacture of bio-oil. Characterization studies

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E-mail address: dewiselvia@feal.unnes.ac.id (D.S. Fardhyanti).

on coconut shells have been done through gravimetric, ultimate, and Thermo Gravimetric (TG) analyses. The mark of this study is to indicate that coconut shell has a high potential to produce fuel fluid by pyrolysis conversion process [16]. Chemical content of coconut shell is cellulose (44%), hemicellulose (21%), lignin (27%) and ash (1%) [17]. Before converted into bio-oil, coconut shell waste and coffee shell need to be treated first. It takes a process before removing the waste of coffee shell into bio-oil which is pyrolysis. Pyrolysis is the process of decomposition of a substance or material that is done at a relatively high temperature. Pyrolysis exists in the form of gas, liquid and solid. The solid left after the process is char (char). Gas and liquid products can be used as a source of bio-oil. Bio-oil can be used as an alternative fuel that is easier to store and distribute than biomass in solid form. Bio-oil made from coconut shell material using slow pyrolysis with pyrolysis temperature 250–300 °C produced a liquid which is very acidic and corrosive. The largest organic component in bio-oil is the liquid derivative of phenol, alcohol, organic acids and carbonyl compounds such as ketones, aldehydes and esters [12]. The nature of the acid is caused by the content of the dominant phenol compounds.

Phenol compounds in addition to causing the smell of smoke in bio-oil, also showed antioxidant activity and corrosivity [13]. Therefore, the use of bio-oil will directly cause some difficulties in the combustion system due to high viscosity, low calorific value, corrosivity and instability [3]. Phenol needs to be extracted from bio-oil to reduce its corrosive properties. In addition, phenol is a compound that has a high economic value because it is expensive and widely used. Phenols have many important uses such as disinfectants, pesticides, explosives and drugs. The method that can be used to extract phenol compounds from bio-oil is by liquid-liquid extraction method using methanol solvent. This has been done by Jadhav [14] on the isolation of phenol compounds from bio-oils from pyrolysis of forest waste and Mantilla [15] on the extraction of phenol compounds from bio-oils from pyrolysis of agricultural waste. Dhalia [16] conducted research on liquid-liquid extraction of phenol compound from bio-oil of pyrolysis in a batch of palm oil empty bunch using methanol solvent. The yield of phenol resulting from this extraction is 40%. In this study, it is expected that the liquid-liquid extraction method can be used to separate the phenol compounds from coconut shell pyrolysis bio-oil. The purpose of liquid extraction is to improve the quality of bio-oil and reduce the corrosivity so when used as fuel, it will cause no damage to the engine.

Biomass pyrolysis converted to bio-oil still has to be done. Bio-oil produced from the pyrolysis process can be used as fuel for engines, turbines and boilers. Further processing by the liquid-liquid extraction process will reduce the corrosive nature of bio-oil and substituted chemicals, in which one of them is phenol. Research on extraction of phenol compounds from bio-oil still very limited. In addition, the thermodynamic modelling study on liquid-liquid equilibrium of the phenol extraction process from bio-oil still needs to be investigated further.

To design extraction columns require much information such as liquid-liquid equilibrium data (liquid-liquid equilibrium). The thermodynamic models such as Equation of State or Activity Coefficient are used to obtain the experimental data of liquid-liquid equilibrium parameters. Optimal parameters can be used to predict liquid-liquid equilibrium data to produce an equilibrium graph that is used to design extraction columns [17]. The study on liquid-liquid equilibrium includes NRTL, e-NRTL, UNIQUAC, and UNIFAC.

Given the importance of thermodynamic modelling in this liquid-liquid extraction process and the price and benefits of phenol in the industrial world, this research becomes an important thing to do. From the equilibrium data, the optimum condition will be obtained where the highest yield of extract is obtained in the extraction process of phenol compound from bio-oil from coconut pyrolysis. Liquid-liquid equilibrium data of the components of the system can be used in the design of the equipment in the separation process.

2. Experimental

2.1. Chemicals

All reagents used here are of AR grade and used as received without further purification. Methanol, acetic acid, and chloroform were purchased from a laboratory (Sigma). The distilled water is available at the laboratory. Coffee shell and coconut shell were obtained from public market in Semarang.

2.2. Equilibrium procedure

Instruments used in this study:

- Shaker bath, product by Memmert NR14, SV1422, Schwabach Germany.
- Gas Chromatography (GC), product by GC 6800 Agilent Technologies, volume: Ratex 800-SM500E.
- Gas Chromatography-Mass Spectrometry (GC-MS), product by GCMS-QP2010S Shimadzu, column: Ratex 800-SM500E.

The material systems and experiment conditions for the equilibrium extraction are summarized in Table 1.

Material system and condition for equilibrium extraction	Quantity
Real volume	30 ml
Approximate amount	Approximate 30 ml
Molar fraction of water in extract	0.2
Ratio of water into	1
Rotation speed	100 rpm
Weight of drying	100 mg
Temperature	200, 111, 121 K

The bio-oil feed solution that contains phenol was extracted using two-stage extraction. The first stage was distilled water as a polar solvent and chloroform a non-polar solvent. Extraction was conducted with the ice bath method at 4 °C, 200 rpm⁻¹ and stirring time for 2 h. The extracts obtained at this stage are used as feeds on the second stage extraction. At the second stage extraction, each extract was extracted using methanol solvent under operating conditions shown in Table 1.

2.3. Analysis and characterization

After equilibrium was attained (90 min), the mixtures were poured into a separating funnel, stirred for an hour and separated into two phases. Then the two phases were weighed. The extract phase and the raffinate phase of each extraction process are analyzed by GC to obtain the concentration of each component. The principal conditions of GC analysis are shown in Table 2.

Condition of analysis using GC	Value
Column (Ratex 800-SM500)	30 m
Carrier gas	12.0 ml min ⁻¹
Carrier gas flow rate	1.0
Injection volume	1.0 μl
Injection temperature	200 °C
Chloroform column	93–973
Flow rate column	0.1
Column	0.1
Detector (FID) temperature	250

3. Results and Discussion

3.1. Bio-oil of coconut

The chromatograms of the GC-MS analysis are shown in Figs. 1 and 2. The results of the GC-MS analysis of the bio-oil showed that they contained more than six chemical compounds such as ethyl ester, phenol, furfural, acetic acid, and others. Table 3 and 4 represent that bio-oil contains more than 6 main components. The percentage (%) of decomposition of the bio-oil analysis by GC-MS was obtained from the percentage area of the peak of the height of the peak in the chromatogram. Using a method of GC-MS, this work presents a composition of the bio-oil generated in the experiment. The total phenolic compounds contained in bio-oil are 47.03% (coconut shell) and 43% (coffee shell). It was caused by the pyrolysis process at 300–350 °C (low temperature carbonization) that produced more substances containing ester and phenolic compounds. The composition of biomass such as hemicellulose, cellulose and lignin will be outlined by phenol as the main component of the bio-oil.

3.2. Equilibrium data as phenol extraction of bio-oil from coconut pyrolysis

In this research the variables studied were temperature, speed of stirring and type of solvent (apexone methanol and aqueous acetic

acid).

The main components of bio-oil produced by pyrolysis of coconut shells	Component	Percentage
1	ethyl ester	17.08
2	phenol	40.02
3	1-ethyl-3-oxo-butan-2-one	3.00
4	furfural	5.03
5	phenol	40.00
7	Methyl glycol	7.02

Table 4

Peak	Component	Percentage
1	ethyl ester	5.8
2	phenol	20.8
3	furfural	0.8
5	phenol	34.8
7	Phenol, 2-methoxy-	7.8
8	Phenol, 2,4-dimethoxy	4.8

Generally, all extracts are rich in phenol but the mole fraction significantly depends on extraction conditions: temperature, speed of stirring and type of solvent. Methanol and acetic acid need to be

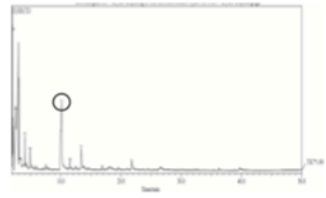


Fig. 1. Analysis result by GC-MS for the bio-oil produced by coconut shell pyrolysis.

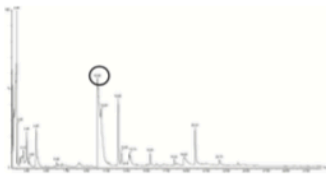


Fig. 2. Analysis result by GC-MS for the bio-oil produced by coffee shell pyrolysis.

mixed with water because their polarities are strong enough to form an immiscible mixture with [17].

The mole fraction of any component in the raffinate phase and the extract phase are constant after 90 min, as the system is assumed to be in equilibrium. The amount of water transferred from the water into the raffinate phase was negligible [17].

In this work, we report experimental data for the phenol extraction from bio-oil produced from coconut shell and the phenol extraction from bio-oil produced from coffee shell.

Table 5 and 6 show the equilibrium of mole fraction extraction data for aqueous acetone and aqueous methanol solvent in the extract phase and raffinate phase.

Table 5

Temperature	Methanol	x_1	x_2	x_3	x_4	x_5	x_6
20K	Coconut shell	0.00	0.00	0.20	0.20	0.01	0.00
	Coffee shell	0.00	0.00	0.20	0.20	0.01	0.00
	Coconut shell	0.00	0.00	0.21	0.21	0.01	0.00
	Coffee shell	0.00	0.00	0.21	0.21	0.01	0.00
	Coconut shell	0.00	0.00	0.22	0.22	0.01	0.00
	Coffee shell	0.00	0.00	0.22	0.22	0.01	0.00
111K	Coconut shell	0.00	0.00	0.20	0.20	0.01	0.00
	Coffee shell	0.00	0.00	0.20	0.20	0.01	0.00
	Coconut shell	0.00	0.00	0.21	0.21	0.01	0.00
	Coffee shell	0.00	0.00	0.21	0.21	0.01	0.00
	Coconut shell	0.00	0.00	0.22	0.22	0.01	0.00
	Coffee shell	0.00	0.00	0.22	0.22	0.01	0.00
121K	Coconut shell	0.00	0.00	0.20	0.20	0.01	0.00
	Coffee shell	0.00	0.00	0.20	0.20	0.01	0.00
	Coconut shell	0.00	0.00	0.21	0.21	0.01	0.00
	Coffee shell	0.00	0.00	0.21	0.21	0.01	0.00
	Coconut shell	0.00	0.00	0.22	0.22	0.01	0.00
	Coffee shell	0.00	0.00	0.22	0.22	0.01	0.00

Table 6

Temperature	Methanol	x_1	x_2	x_3	x_4	x_5	x_6
20K	Coconut shell	0.00	0.247	0.305	0.305	0.420	0.022
	Coffee shell	0.00	0.239	0.294	0.291	0.327	0.027
	Coconut shell	0.00	0.250	0.305	0.305	0.420	0.022
	Coffee shell	0.00	0.247	0.275	0.275	0.343	0.027
	Coconut shell	0.00	0.264	0.315	0.315	0.422	0.022
	Coffee shell	0.00	0.259	0.271	0.271	0.322	0.027
111K	Coconut shell	0.00	0.244	0.297	0.298	0.328	0.028
	Coffee shell	0.00	0.234	0.286	0.285	0.329	0.028
	Coconut shell	0.00	0.251	0.304	0.305	0.421	0.022
	Coffee shell	0.00	0.249	0.287	0.285	0.324	0.028
	Coconut shell	0.00	0.267	0.318	0.318	0.422	0.022
	Coffee shell	0.00	0.254	0.290	0.289	0.324	0.028
121K	Coconut shell	0.00	0.234	0.286	0.285	0.324	0.028
	Coffee shell	0.00	0.224	0.276	0.275	0.311	0.027
	Coconut shell	0.00	0.241	0.297	0.295	0.321	0.027
	Coffee shell	0.00	0.234	0.286	0.285	0.324	0.028
	Coconut shell	0.00	0.254	0.304	0.304	0.421	0.022
	Coffee shell	0.00	0.241	0.297	0.295	0.321	0.027

3.3. Liquid-liquid equilibrium for the phenol extraction process using UNIFAC and NRTL model

The UNIFAC and NRTL models were used to correlate the experimental data obtained for the systems under study. The UNIFAC equation and

NRTL equation [18] were used to correlate experiment of phase equilibrium data. The activity coefficient models applied in this study are as follows:

a) UNIFAC [18]

UNIFAC is a UNIFAC Functional group Activity Coefficient model based on the UNIFAC equation by using functional groups of molecules contained in the molecule to calculate the activity coefficient. The UNIFAC modeling is a continuation of the UNIFAC modeling development. This model has the advantage of performing phase equilibrium calculation in the absence of experimental data. This thermodynamic model is used to predict or estimation of non-electrolyte activity in ideal mixtures. The UNIFAC model calculates the activity coefficients by using functional groups present in the molecules that from the liquid mixture if the value of the group contribution coefficient has a value close to zero, this indicates that the component is in the ideal state in the system [18]. The correlation in the UNIFAC model is used in order to reduce the number of binary interaction measured to predict the state of the system. The correlation in the UNIFAC model is shown as follows:

$$\ln \gamma_i = -\ln \gamma_i^* + \ln \gamma_i^* \quad (1)$$

$$\ln \gamma_i^* = 1 - \ln \gamma_i^* - \ln \gamma_i^* \left(\frac{1}{\gamma_i^*} - \frac{1}{\gamma_i^*} \right) \quad (2)$$

$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (3)$$

$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (4)$$

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$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (20)$$

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$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (35)$$

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$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (39)$$

$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (40)$$

$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (41)$$

$$\ln \gamma_i^* = \ln \gamma_i^* - \ln \gamma_i^* \quad (42)$$

where:

γ_i^* : mole Gibbs energy

R : gas constant

T : temperature

x : liquid phase mole fraction

y : component of 1, 2, 3, ...

α : non-random parameter for NRTL

β : GC component parameter

γ : activity coefficient

τ : empirical constant for NRTL

Table 7 and 8 show the value of activity coefficient on phenol extraction using methanol solvent obtained from calculation with UNIFAC and NRTL modeling, and the coefficient value of activity on each component shows that methanol and acetic acid component in the extract phase is in ideal condition because the value of activity coefficient obtained is close to one. The activity coefficient value of phenol and water in the extract phase is not close to one, it means that phenol and water components are far from ideal conditions for the system. The ideal conditions of the components in the system show that the component is dominant, and the properties of the mixture are close to the dominant component properties either in the extract or raffinate phase. In the raffinate phase, it is known that the bio-oil has a coefficient of activity close to one, indicating that the bio-oil is in ideal conditions for

Table 6
The activity coefficients were calculated by NRTL model

Solution	Temperature	Solvent	Solute					
			γ_1^E	γ_2^E	γ_3^E	γ_4^E	γ_5^E	
Methanol	20 K	Cocaine shell	0.077	0.086	0.261	0.085	0.236	2.027
		Cocaine	0.064	0.062	0.127	0.061	0.228	2.027
		Coffee shell	0.063	0.066	0.267	0.131	0.146	2.045
	100 K	Cocaine shell	0.152	1.063	0.269	0.081	0.166	1.803
		Cocaine	0.157	1.065	0.276	0.081	0.169	1.823
		Coffee shell	0.021	1.067	0.491	0.087	0.260	0.906
100 K	Cocaine shell	4.47×10^{-4}	1.065	0.010	8.4×10^{-4}	0.228	1.929	
		1.71×10^{-4}	1.066	0.056	1.5×10^{-4}	0.260	0.906	
		1.80×10^{-4}	1.025	0.059	4.9×10^{-4}	0.247	0.889	
	Cocaine	0.065	0.067	0.080	0.063	0.260	0.906	
		0.065	0.067	0.080	0.063	0.260	0.912	
		0.067	1.068	0.130	0.067	0.260	1.028	
200 K	Cocaine shell	7.92×10^{-4}	1.110	0.010	3.4×10^{-4}	0.116	1.028	
		3.46×10^{-4}	1.068	0.056	4.1×10^{-4}	0.132	1.086	
		6.84×10^{-4}	1.122	0.054	3.5×10^{-4}	0.116	0.927	
	Cocaine	0.065	1.323	0.054	0.064	0.107	1.022	
		0.066	1.422	0.057	0.067	0.115	1.022	
		0.066	1.389	0.055	0.103	0.115	1.022	
Acetic acid	20 K	Cocaine shell	0.046	1.299	0.218	0.042	0.293	1.987
		Cocaine	0.059	1.211	0.210	0.015	0.280	1.205
		Coffee shell	0.056	1.269	0.223	9.5×10^{-4}	0.280	1.478
	100 K	Cocaine shell	0.146	1.488	0.067	0.057	0.236	2.014
		Cocaine	0.125	1.405	0.075	0.059	0.247	2.004
		Coffee shell	0.128	1.761	0.102	0.078	0.228	2.002
100 K	Cocaine shell	0.062	2.043	0.067	0.060	0.126	0.905	
		0.063	2.052	0.070	0.059	0.131	0.906	
		0.063	1.956	0.069	0.058	0.117	1.000	
	Cocaine	0.069	1.824	0.026	0.026	0.210	1.118	
		0.069	1.762	0.023	0.022	0.209	1.119	
		0.069	1.824	0.026	0.025	0.210	1.118	
200 K	Cocaine shell	0.115	1.546	0.037	0.035	0.208	1.129	
		0.126	1.689	0.036	0.041	0.210	1.016	
		0.240	1.890	0.030	8.9×10^{-4}	0.280	1.023	
	Cocaine	0.065	1.686	0.031	0.031	0.207	1.206	
		0.065	2.025	0.037	0.034	0.238	1.017	
		0.065	2.068	0.039	0.034	0.228	1.229	

Note: γ_1^E = mole fraction of 1 component in extract phase
 γ_2^E = mole fraction of solvent in extract phase
 γ_3^E = mole fraction of water in extract phase
 γ_4^E = mole fraction of component in raffinate phase
 γ_5^E = mole fraction of solvent in raffinate phase
 γ_6^E = mole fraction of component in raffinate phase
 γ_7^E = mole fraction of solvent in raffinate phase

the system, while the phenol and solvent (acetic acid or methanol) are far from ideal conditions. So non-ideality studies are very important for these systems [10].
 The mean of correlation coefficient liquid-liquid equilibrium data by using UNIQUAC and NRTL modeling give good agreement between experimental data and calculation. This is shown by the value of SSE (Sum of the Square of the Error), which is very small. The SSE values in UNIQUAC and NRTL modeling can be seen in Table 5. The average of SSE is 0.015–0.554.

Table 9
Sum of the Square of the Error (SSE)

Model	Temperature	SSE	
		Methanol	Acetic acid
UNIQUAC	200	0.040	0.122
	100	0.016	0.062
	20	0.016	0.084
NRTL	200	0.006	0.054
	100	0.016	0.054
	20	0.016	0.077

Based on Table 9, it can be seen that extraction with methanol solvent for the NRTL model has the smallest SSE value compared to the extraction with acetic acid solvent for the UNIQUAC model.

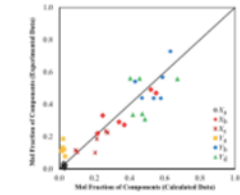


Fig. 3 Comparison of calculated and experimental mole fractions of the phenol extract using methanol by UNIQUAC model at 20 K.
 γ_1^E = mole fraction of component in extract phase
 γ_2^E = mole fraction of solvent in extract phase
 γ_3^E = mole fraction of water in extract phase
 γ_4^E = mole fraction of component in raffinate phase
 γ_5^E = mole fraction of solvent in raffinate phase

Table 6
The activity coefficients were calculated by NRTL model

Solution	Temperature	Solvent	Solute					
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		Coffee shell	0.063	0.066	0.267	0.131	0.146	2.045
	100 K	Cocaine shell	0.152	1.063	0.269	0.081	0.166	1.803
		Cocaine	0.157	1.065	0.276	0.081	0.169	1.823
		Coffee shell	0.021	1.067	0.491	0.087	0.260	0.906
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		1.71×10^{-4}	1.066	0.056	1.5×10^{-4}	0.260	0.906	
		1.80×10^{-4}	1.025	0.059	4.9×10^{-4}	0.247	0.889	
	Cocaine	0.065	0.067	0.080	0.063	0.260	0.906	
		0.065	0.067	0.080	0.063	0.260	0.912	
		0.067	1.068	0.130	0.067	0.260	1.028	
200 K	Cocaine shell	7.92×10^{-4}	1.110	0.010	3.4×10^{-4}	0.116	1.028	
		3.46×10^{-4}	1.068	0.056	4.1×10^{-4}	0.132	1.086	
		6.84×10^{-4}	1.122	0.054	3.5×10^{-4}	0.116	0.927	
	Cocaine	0.065	1.323	0.054	0.064	0.107	1.022	
		0.066	1.422	0.057	0.067	0.115	1.022	
		0.066	1.389	0.055	0.103	0.115	1.022	
Acetic acid	20 K	Cocaine shell	0.046	1.299	0.218	0.042	0.293	1.987
		Cocaine	0.059	1.211	0.210	0.015	0.280	1.205
		Coffee shell	0.056	1.269	0.223	9.5×10^{-4}	0.280	1.478
	100 K	Cocaine shell	0.146	1.488	0.067	0.057	0.236	2.014
		Cocaine	0.125	1.405	0.075	0.059	0.247	2.004
		Coffee shell	0.128	1.761	0.102	0.078	0.228	2.002
100 K	Cocaine shell	0.062	2.043	0.067	0.060	0.126	0.905	
		0.063	2.052	0.070	0.059	0.131	0.906	
		0.063	1.956	0.069	0.058	0.117	1.000	
	Cocaine	0.069	1.824	0.026	0.026	0.210	1.118	
		0.069	1.762	0.023	0.022	0.209	1.119	
		0.069	1.824	0.026	0.025	0.210	1.118	
200 K	Cocaine shell	0.115	1.546	0.037	0.035	0.208	1.129	
		0.126	1.689	0.036	0.041	0.210	1.016	
		0.240	1.890	0.030	8.9×10^{-4}	0.280	1.023	
	Cocaine	0.065	1.686	0.031	0.031	0.207	1.206	
		0.065	2.025	0.037	0.034	0.238	1.017	
		0.065	2.068	0.039	0.034	0.228	1.229	

Note: γ_1^E = mole fraction of 1 component in extract phase
 γ_2^E = mole fraction of solvent in extract phase
 γ_3^E = mole fraction of water in extract phase
 γ_4^E = mole fraction of component in raffinate phase
 γ_5^E = mole fraction of solvent in raffinate phase

the system, while the phenol and solvent (acetic acid or methanol) are far from ideal conditions. So non-ideality studies are very important for these systems [10].
 The mean of correlation coefficient liquid-liquid equilibrium data by using UNIQUAC and NRTL modeling give good agreement between experimental data and calculation. This is shown by the value of SSE (Sum of the Square of the Error), which is very small. The SSE values in UNIQUAC and NRTL modeling can be seen in Table 5. The average of SSE is 0.015–0.554.

Table 9
Sum of the Square of the Error (SSE)

Model	Temperature	SSE	
		Methanol	Acetic acid
UNIQUAC	200	0.040	0.122
	100	0.016	0.062
	20	0.016	0.084
NRTL	200	0.006	0.054
	100	0.016	0.054
	20	0.016	0.077

Based on Table 9, it can be seen that extraction with methanol solvent for the NRTL model has the smallest SSE value compared to the extraction with acetic acid solvent for the UNIQUAC model.

Table 6
The activity coefficients were calculated by NRTL model

Solution	Temperature	Solvent	Solute					
			γ_1^E	γ_2^E	γ_3^E	γ_4^E	γ_5^E	
Methanol	20 K	Cocaine shell	0.077	0.086	0.261	0.085	0.236	2.027
		Cocaine	0.064	0.062	0.127	0.061	0.228	2.027
		Coffee shell	0.063	0.066	0.267	0.131	0.146	2.045
	100 K	Cocaine shell	0.152	1.063	0.269	0.081	0.166	1.803
		Cocaine	0.157	1.065	0.276	0.081	0.169	1.823
		Coffee shell	0.021	1.067	0.491	0.087	0.260	0.906
100 K	Cocaine shell	4.47×10^{-4}	1.065	0.010	8.4×10^{-4}	0.228	1.929	
		1.71×10^{-4}	1.066	0.056	1.5×10^{-4}	0.260	0.906	
		1.80×10^{-4}	1.025	0.059	4.9×10^{-4}	0.247	0.889	
	Cocaine	0.065	0.067	0.080	0.063	0.260	0.906	
		0.065	0.067	0.080	0.063	0.260	0.912	
		0.067	1.068	0.130	0.067	0.260	1.028	
200 K	Cocaine shell	7.92×10^{-4}	1.110	0.010	3.4×10^{-4}	0.116	1.028	
		3.46×10^{-4}	1.068	0.056	4.1×10^{-4}	0.132	1.086	
		6.84×10^{-4}	1.122	0.054	3.5×10^{-4}	0.116	0.927	
	Cocaine	0.065	1.323	0.054	0.064	0.107	1.022	
		0.066	1.422	0.057	0.067	0.115	1.022	
		0.066	1.389	0.055	0.103	0.115	1.022	
Acetic acid	20 K	Cocaine shell	0.046	1.299	0.218	0.042	0.293	1.987
		Cocaine	0.059	1.211	0.210	0.015	0.280	1.205
		Coffee shell	0.056	1.269	0.223	9.5×10^{-4}	0.280	1.478
	100 K	Cocaine shell	0.146	1.488	0.067	0.057	0.236	2.014
		Cocaine	0.125	1.405	0.075	0.059	0.247	2.004
		Coffee shell	0.128	1.761	0.102	0.078	0.228	2.002
100 K	Cocaine shell	0.062	2.043	0.067	0.060	0.126	0.905	
		0.063	2.052	0.070	0.059	0.131	0.906	
		0.063	1.956	0.069	0.058	0.117	1.000	
	Cocaine	0.069	1.824	0.026	0.026	0.210	1.118	
		0.069	1.762	0.023	0.022	0.209	1.119	
		0.069	1.824	0.026	0.025	0.210	1.118	
200 K	Cocaine shell	0.115	1.546	0.037	0.035	0.208	1.129	
		0.126	1.689	0.036	0.041	0.210	1.016	
		0.240	1.890	0.030	8.9×10^{-4}	0.280	1.023	
	Cocaine	0.065	1.686	0.031	0.031	0.207	1.206	
		0.065	2.025	0.037	0.034	0.238	1.017	
		0.065	2.068	0.039	0.034	0.228	1.229	

Note: γ_1^E = mole fraction of 1 component in extract phase
 γ_2^E = mole fraction of solvent in extract phase
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